

SO_DYFAMED Time Series - 1991-> ...

JC. MARTY : head of mission and project leader

PIGMENTS HPLC : [JC. MARTY](#)

[METHOD](#) | [PUBLICATIONS](#) | [FIGURES](#)

(get data set in excel file format : )

METHOD FOR HPLP PIGMENTS ANALYSIS

Water for pigment analysis (2 liters) was filtrated on 25 mm Whatman GF/F glass fiber filter, frozen and analyzed by HPLC within 3 months. Filters were ground and sonicated in 3-ml methanol under dim light conditions. The method used until 1993 was derived from that of Mantoura *et al.* (1994 a,b). The general procedure for HPLC pigment analysis, identification and quantification has been described by Mantoura *et al.* (1994 a,b). With the separation system used (RP-C18), a partial resolution of divinyl-chlorophyll *a* (DV Chl *a*) from chlorophyll *a* (Chl *a*) has been achieved. For samples from 1994 and later, the separation method (RP-C8) is described in Vidussi *et al.* (1996), and the resolution between DV Chl *a* and Chl *a* has been improved.

The continuity of the set of data was obtained by the utilization of an internal standard (β -apocaro-*l*-xanthin) in each sample in the extraction solvent. The possible effect of the change of analytical method on analyzing the same samples with the two procedures. The agreement between the two methods was of the same order than the agreement between 2 analyses of the same sample using the same method. Attention was given to the quantification of DV Chl *a*, which is fully resolved from Chl *a* in the experiment. Although partial, the separation of these two compounds in the first phase of our experiment for a good matching of the data from the two methods (equivalent to other pigment concentrations of DV Chl *a* (below 5 ng l⁻¹) not detected in the first method.

Results are reported in terms of Chl *a*, divinyl-chlorophyll *a* (DV Chl *a*) and Total Chl *a* (TChl *a* = Chlorophyll *b* (Chl *b*) and divinyl-chlorophyll *b* (DV Chl *b*) not resolved with the first separation method, partially resolved by the new one, are presented together as TChl *b*. Lutein and zeaxanthin were separated using the method of Vidussi *et al.* (1996), but data are presented as the sum of the two compounds. Zeaxanthin was only occasionally detected and always at very low levels with respect to zeaxanthin. The zeaxanthin can be considered as essentially zeaxanthin.

Chlorophylls and carotenoids were detected and quantified by absorbance at 440 nm. Identification was performed by comparison of on-line collected absorption spectra with those of a library of pigments from standards and reference cultures obtained from the Villefranche sur mer culture collection. Carotenoids used for the calibration of the HPLC [peridinin (peri), alloxanthin (allo), fucoxanthin (zea), 19'-hexanoyloxyfucoxanthin (19'HF), 19'-butanoyloxyfucoxanthin (19'BF)] were provided as part of a JGOFS intercalibration exercise. Chlorophyll *a* and chlorophyll *b* were from Sigma. Array detection was achieved on selected samples until 1993 (Waters 991) and on all samples from 1994 (Waters 1100).

A range of phytoplankton pigments has been detected, in order to characterize different phytoplankton communities.

recent review of taxonomic pigments can be found in Jeffrey (1997). Divinyl-chlorophyll *a* is the marker of prochlorophytes whereas Chl *a* is the universal descriptor of other phytoplankton taxa. Chl *b* characterizes diatoms and peridinin (peri) dinoflagellates. Nano- and pico-flagellates contain pigments characterized by 19'-hexanoyloxyfucoxanthin (19'HF, prymnesiophytes) and by 19'-butadienylfucoxanthin (19'BF, chrysophytes and pelagophytes). Zeaxanthin (Zea) is the marker of cyanobacteria but prochlorophytes.

All data are available through the DYFAMED Observatory data base vlfr.fr/jgofs2/sodyf/home.htm.

Contour maps were obtained using Surfer program (Golden software Inc.) and Kriging method.

BIBLIOGRAPHY

Claustre, H., Kerhervé, P., Marty, J.C., Prieur, L., Videau, C., Hecq, J.H., 1994. Phytoplankton distribution and structure in the presence of a geostrophic front: ecological and biogeochemical implications. *Journal of Marine Research* 52, 101-115.

Claustre, H., Kerhervé, P., Marty, J.C., Prieur, L., 1994. Phytoplankton photoadaptation in relation to physical processes. *Journal of Marine Systems* 5, 251-265.

Jeffrey, S.W., 1997. Application of pigment methods to oceanography. In: Jeffrey, S.W., Mantoura S.W. (Eds.), *Phytoplankton pigments in oceanography*. UNESCO, Paris, pp. 127-178.

Mantoura, R.F.C., Llewellyn, C.A., 1983. The rapid determination of algal chlorophyll and carotenoids and their breakdown products in natural waters by reverse-phase high-performance liquid chromatography. *Chimica Acta* 151, 293-314.

Vidussi, F., Claustre, H., Bustillos-Guzman, J., Cailliau, C., Marty, J.C., 1996. Determination of carotenoids of marine phytoplankton : separation of chlorophyll *a* from divinyl-chlorophyll *a* and lutein. *Journal of Plankton Research* 18, 2377-2382.

FIGURES





