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Natural spatial variability of depositional conditions, biogeochemical processes and element fluxes in sediments of the eastern Clarion-Clipperton Zone, Pacific Ocean

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ABSTRACT

The manganese nodule belt within the Clarion and Clipperton Fracture Zones (CCZ) in the abyssal NE Pacific Ocean is characterized by numerous seamounts, low organic matter (OM) depositional fluxes and meter-scale oxygen penetration depths (OPD) into the sediment. The region hosts contract areas for the exploration of polymetallic nodules and Areas of Particular Environmental Interest (APEI) as protected areas. In order to assess the impact of potential mining on these deep-sea sediments and ecosystems, a thorough determination of the natural spatial variability of depositional and geochemical conditions as well as biogeochemical processes and element fluxes in the different exploration areas is required.

Here, we present a comparative study on (1) sedimentation rates and bioturbation depths, (2) redox zonation of the sediments and element fluxes as well as (3) rates and pathways of biogeochemical reactions at six sites in the eastern CCZ. The sites are located in four European contract areas and in the APEI3. Our results demonstrate that the natural spatial variability of depositional and (bio)geochemical conditions in this deep-sea sedimentary environment is much larger than previously thought. We found that the OPD varies between 1 and 4.5 m, while the sediments at two sites are oxic throughout the sampled interval (7.5 m depth). Below the OPD, manganese and nitrate reduction occur concurrently in the suboxic zone with pore-water Mn^{2+} concentrations of up to 25 μM . The thickness of the suboxic zone extends over depth intervals of less than 3 m to more than 8 m. Our data and the applied transport-reaction model suggest that the extension of the oxic and suboxic zones is ultimately determined by the (1) low flux of particulate organic carbon (POC) of 1–2 $\text{mg C}_{\text{org}} \text{m}^{-2} \text{d}^{-1}$ to the seafloor, (2) low sedimentation rates between 0.2 and 1.15 cm kyr^{-1} and (3) oxidation of pore-water Mn^{2+} at depth. The diagenetic model reveals that aerobic respiration is the main biogeochemical process driving OM degradation. Due to very low POC fluxes of 1 $\text{mg C}_{\text{org}} \text{m}^{-2} \text{d}^{-1}$ to the seafloor at the site investigated in the protected APEI3 area, respiration rates are twofold lower than at the other study sites. Thus, the APEI3 site does not represent the (bio)geochemical conditions that prevail in the other investigated sites located in the European contract areas. Lateral variations in surface water productivity are generally reflected in the POC fluxes to the seafloor across the various areas but deviate from this trend at two of the study sites. We suggest that the observed spatial variations in depositional and (bio)geochemical conditions result from differences in the degree of degradation of OM in the water column and heterogeneous sedimentation patterns caused by the interaction of bottom water currents with seafloor topography.

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1. Introduction

Vast deep-sea regions of the open Pacific Ocean are characterized by low fluxes of organic carbon to the seafloor (Lutz et al., 2007). The discovery of high abundances of polymetallic nodules in these carbon-starved environments in the equatorial Pacific Ocean during the *HMS Challenger* expedition (Murray and Renard, 1891) has driven the global economic interest in deep-sea mining (Mero, 1965) and has triggered several comprehensive studies in the area of the Clarion-Clipperton Zone (CCZ) (e.g., Chun, 1908; Bischoff and Piper, 1979; Bender, 1983; von Stackelberg and Beiersdorf, 1987; Lodge et al., 2014). Most geochemical studies on CCZ sediments have focused on polymetallic manganese nodules, more precisely their chemical and mineralogical composition as well as pathways of formation (e.g., Calvert and Price, 1977; Jeong et al., 1994; Wegorzewski and Kuhn, 2014). Only a few studies that combine pore-water and sediment geochemistry have been performed on sediments of the CCZ (e.g., Bischoff and Piper, 1979; Jahnke et al., 1982; Müller et al., 1988; Mewes et al., 2014; Mogollón et al., 2016). The upper few centimeters of the sediments in the CCZ are generally characterized by organic carbon (OC) contents of < 0.5 wt% (e.g., Arrhenius, 1952; Heath et al., 1977; Khripunoff et al., 2006). Below 30 cm, a residual OC fraction of ~ 0.1 wt% remains in the deeply buried sediments (Müller and Mangini, 1980; Müller et al., 1988; Mewes et al., 2014; Mogollón et al., 2016). Müller and Mangini (1980) have reported a sedimentation rate of 0.15–0.4 cm kyr⁻¹ for the western CCZ. Slightly higher rates between 0.35 and 0.6 cm kyr⁻¹ were determined by Mewes et al. (2014) for the eastern CCZ. In the framework of the MANOP (Manganese Nodule Project) study, sediments and associated pore water underlying the Pacific equatorial upwelling area between 0° and 10°N were analyzed (e.g., Emerson et al., 1980; Klinkhammer, 1980). The relatively high POC flux to the seafloor in this area (Lutz et al., 2007) causes a compressed redox zonation in the sediments with the occurrence of Mn²⁺ in the absence of oxygen and sulfide in the pore water after nitrate is consumed below 10 cm (Emerson et al., 1980; Klinkhammer, 1980; Jahnke et al., 1982). The study by Røy et al. (2012) was performed along a sampling transect at 0°N in the eastern Pacific Ocean and shows that the oxygen penetration depth (OPD) is generally 10 cm. Oxygen depletion in combination with the absence of sulfate reduction allows manganese reduction to extend over sediment intervals of up to 100 m (D'Hondt et al., 2004). Studies performed by Mewes et al. (2014) and Mogollón et al. (2016) in the German contract area “East” for the exploration of polymetallic nodules show that oxygen typically penetrates 1.8–3 m into the sediments between 11°–12°N and 117°–120°W. Below the oxic zone, manganese and nitrate reduction occur concurrently over depth intervals between 6 m to more than 12 m depth. In contrast, sediments underlying the carbon-starved waters of the North Pacific Gyre (NPG) are deeply oxygenated at least 30 m below the seafloor due to low respiration rates (Røy et al., 2012).

Considering that abyssal benthic communities are limited by low carbon export from the euphotic zone (e.g., Smith et al., 2008), biodiversity in the CCZ is surprisingly high (e.g., Glover et al., 2002). Vanreusel et al. (2016) have recently performed a comparative study of benthic faunal composition and densities in five areas in the CCZ over a distance of approximately 1300 km. This biological study revealed (1) lowest densities of both sessile and mobile fauna in the area with lowest POC fluxes to the seafloor, (2) a strong dependency of local biodiversity on manganese nodule abundance and (3) a reduction of the mobile fauna by at least 50% in areas where controlled anthropogenic disturbances were created 37 and 20 years ago. As these simulated deep-sea mining experiments cause the removal or alteration of top sediment layers, and the formation of large sediment plumes in the water column, the recovery of benthic communities after anthropogenic impacts is expected to be very slow (e.g., Miljutin et al., 2011; Jones et al., 2017; Boetius and Haeckel, 2018).

Due to the economic interest in the exploitation of deep-sea

manganese nodules, the International Seabed Authority (ISA) has adopted an Environmental Management Plan (EMP) for the CCZ, which includes temporal contracts for the exploration of manganese nodules (ISA, 2010; Lodge et al., 2014; Madureira et al., 2016). In addition to this, nine areas have been designated for the conservation of natural resources, which are excluded from any mining activities and declared as “Areas of Particular Environmental Interest” (APEI). For the further development of an efficient EMP, Lodge et al. (2014) emphasize the necessity of environmental baseline studies including the determination of chemical parameters before and after anthropogenic disturbances.

In order to assess the natural spatial variability of geochemical conditions, biogeochemical processes, and element fluxes in the CCZ as needed for such baseline studies, we present a comparative study on (1) sedimentation rates and bioturbation depths, (2) redox zonation of sediments and oxygen fluxes, and (3) rates and pathways of biogeochemical reactions driving the degradation of organic matter in these deep-sea deposits. We have studied sediments of four European contract areas including the German BGR area “East” (Bundesanstalt für Geowissenschaften und Rohstoffe), the area of the eastern European consortium IOM (InterOceanMetal), the Belgian GSR area (Global Sea Mineral Resources NV), the French IFREMER area (Institut Français de Recherche pour l'Exploitation de la Mer) and one of the nine APEIs which is located north of the CCZ and referred to as the APEI3. Our work includes *ex situ* oxygen measurements, comprehensive pore-water and solid-phase analyses on the upper 10 m of sediment and the application of a one-dimensional steady-state reaction transport model.

2. Geological and oceanographic setting

The CCZ comprises an area of about 6 million km² in the equatorial Pacific Ocean defined by two major transform faults, the Clarion Fracture Zone in the north and the Clipperton Fracture Zone in the south (Halbach et al., 1988). The fracture zones are formed at the East Pacific Rise, stretch perpendicular to the spreading center, and enclose a vast seafloor covered by numerous seamounts and NNE-SSW oriented horst and graben structures (Johnson, 1972; ISA, 2010). The abyssal deep-sea pelagic sediments at 4–5 km water depth between 116°–155°W and 5°–15°N are dominated by biogenic sediments, notably siliceous oozes (Berger, 1974). The CCZ sedimentary records commonly have hiatuses caused by (1) erosion and redeposition of sediment elsewhere or (2) nondeposition (Johnson, 1972; Craig, 1979; von Stackelberg and Beiersdorf, 1991; Mewes et al., 2014). Average deep-water flow velocities of < 10 cm s⁻¹ refute significant contemporary erosion, but distinct hiatuses in the Eocene to Quaternary sediments may have resulted from relatively higher current velocities (Craig, 1979; Theyer et al., 1985; von Stackelberg and Beiersdorf, 1987).

Average particulate organic carbon (POC) fluxes to the seafloor in the eastern CCZ are 1.5–1.8 mg C_{org} m⁻² d⁻¹ between 10° and 15°N and decline to 1.3 mg C_{org} m⁻² d⁻¹ north of 15°N (Lutz et al., 2007; Fig. 1a). As a consequence of microbial respiration in the water column and weak ocean ventilation, a pronounced oxygen minimum zone (OMZ) persists in the eastern equatorial Pacific (e.g., Wishner et al., 1995; Kalvelage et al., 2015). Oxygen measurements throughout the water column in the CCZ show a well-oxygenated upper mixed layer with a sharp oxycline particularly in the BGR and IOM areas (Martínez Arbizu and Haeckel, 2015). Below the oxycline, the OMZ extends over 100–1000 m and 100–800 m, respectively, with concentrations below 3 μM. The GSR and IFREMER areas show similar extents of the OMZ, however, lowest oxygen concentrations range between 3 and 16 μM. In the APEI3, the OMZ is located at 300–900 m water depth with minimum oxygen concentrations of 6–16 μM (Martínez Arbizu and Haeckel, 2015).

3. Material and methods

As part of the BMBF-EU JPI Oceans pilot action “Ecological Aspects

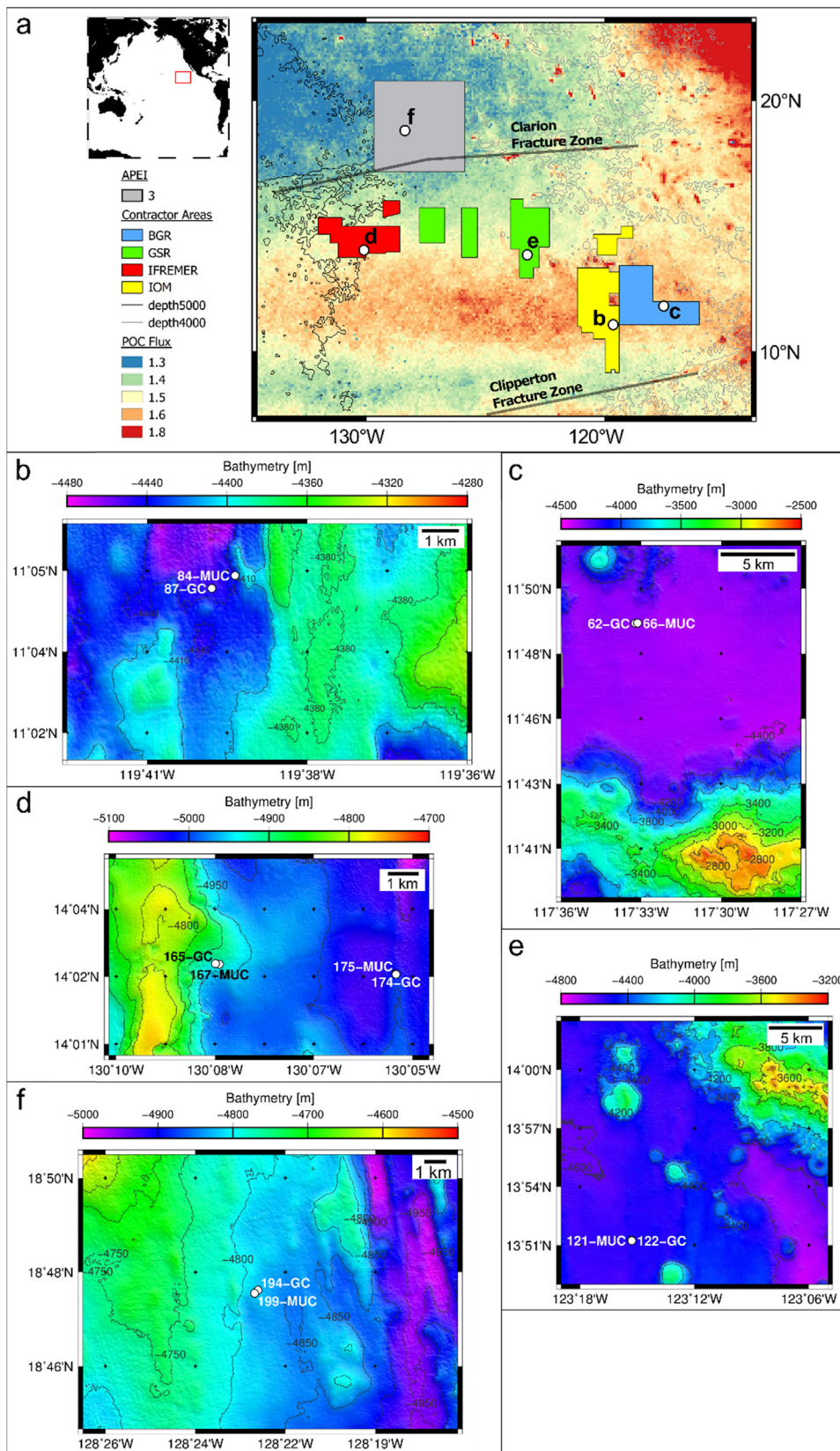


Fig. 1. Maps of the study area during RV SONNE cruise SO239 showing (a) the four investigated exploration areas in the CCZ, the APEI3 and the sampling stations (white circles). The background colors indicate the estimated upper limit in POC flux [mg C_{org} m⁻² d⁻¹] to the seafloor by Lutz et al. (2007) (modified after Vanreusel et al., 2016). Detailed hydro-acoustic maps created with the multibeam system EM122 (Martínez Arbizu and Haeckel, 2015; Greinert, 2016) show the locations of the sampling stations with b: IOM; c: BGR; d: IFREMER; e: GSR; f: APEI3. The maps were produced with GMT5 (Wessel et al., 2013). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

of Deep-Sea Mining (MiningImpact)™ sediment cores were taken at five sites in the CCZ during RV SONNE cruise SO239 in March/April 2015 (Fig. 1; Martínez Arbizu and Haeckel, 2015). For the recovery of sediment cores, two different sampling devices were deployed in all investigated areas. A multiple corer (MUC) equipped with twelve 60 cm

long tubes with an inner diameter of 9.4 cm was used for the retrieval of undisturbed surface sediments. For the recovery of long sediment cores of up to 10 m length a gravity corer (GC) with a 12 cm wide plastic liner was deployed (Table 1).

Table 1

MUC and GC cores investigated in this study including information on geographic position, water depth and core length. Nodule coverage shows the density and size of nodules recovered from the sediment surface at the box corer (BC) station nearest to MUC and GC locations. Nodule sampling area is 0.25 m², the length of scale (■) corresponds to 10 cm.

Area	Station SO239-	Device	Latitude [N]	Longitude [W]	Water depth [m]	Core length [cm]	Nodule coverage
BGR	60	BC	11°48.46'	117°33.02'	4324.5		
BGR	62	GC	11°49.12'	117°33.22'	4312.2	900	
BGR	66	MUC	11°49.13'	117°33.13'	4314.8	36	
IOM	84	MUC	11°4.73'	119°39.48'	4430.8	43	
IOM	87	GC	11°4.54'	119°39.83'	4436	930	
IOM	89	BC	11°4.55'	119°39.65'	4436.5		
GSR	121	MUC	13°51.25'	123°15.3'	4517.7	30	
GSR	122	GC	13°51.23'	123°15.29'	4517.7	740	
GSR	128	BC	13°51.10'	123°15.12'	4510.7		
IFRE-1	165	GC	14°2.63'	130°8.39'	4922.7	927	
IFRE-1	167	MUC	14°2.62'	130°8.32'	4918.8	29	
IFRE-1	180	BC	14°2.50'	130°8.18'	4936.4		
IFRE-2	174	GC	14°2.44'	130°5.1'	5008	734	no nodules
IFRE-2	175	MUC	14°2.45'	130°5.11'	5005.5	36	
APEI3	194	GC	18°47.54'	128°22.33'	4815.5	576	
APEI3	195	BC	18°47.75'	128°21.73'	4833.4		
APEI3	199	MUC	18°47.46'	128°22.42'	4816.6	32	

3.1. Pore-water and sediment sampling

Immediately after core recovery, sediment cores were transferred into the cold room of the RV SONNE at a temperature of ~ 4 °C. Two MUC cores were separately used for (1) oxygen measurements and subsequent solid-phase sampling and (2) the retrieval of bottom water and pore water by means of rhizons with an average pore size of 0.1 μm (Seeberg-Elverfeldt et al., 2005). Pore water was sampled at intervals of 1 cm in the upper 10 cm and at 2 cm below. For solid-phase investigations the sample interval for the MUC cores was 1 cm.

The GCs were cut into 1 m segments on deck and stored in the cold room for at least 12 h in order to allow temperature equilibration of the sediments before oxygen measurements were performed. After the oxygen measurements, the segments were split into two halves from which the 'working half' was used for pore-water and sediment

sampling while the other half was kept undisturbed and archived. Pore-water and sediment from the GCs were sampled every 20 cm.

During the pore-water sampling by rhizons, the first mL of extracted pore water was discarded in order to avoid any dilution or oxidation. Sample aliquots of typically 2 mL for nitrate (NO₃) were stored in amber vials sealed with a PTFE septum-bearing lid at - 20 °C. For the analyses of further dissolved pore-water constituents, aliquots of the remaining pore-water samples were diluted 1:10 and acidified with 0.145 M sub-boiling distilled HNO₃ and stored at 4 °C in Zinsser vials. All sediment samples were taken using either cut-off syringes with a volume of about 12 mL or with a plastic spatula and stored at - 20 °C in plastic vials. Sediment samples from suboxic intervals of the cores were stored in argon-flushed gas-tight glass bottles at - 20 °C until further analysis.

3.2. Ex situ oxygen measurements

Oxygen concentrations in the sediment were determined using amperometric Clark-type oxygen sensors with an internal reference and equipped with a guard cathode (Revsbech, 1989) according to the procedure described by Ziebis et al. (2012) and Mewes et al. (2014). The electrodes (Unisense, Denmark) are made of glass with a 6 cm long tip that was inserted into a hyperdermic needle (diameter 1.1 mm, length 50 mm) and had a response time shorter than 10 s. Signals were amplified and transformed to mV by a picoamperemeter, digitalized by an analogue/digital converter (ADC 216, Unisense, Denmark) and recorded using the software PROFIX (Unisense, Denmark). Measurements were recorded at each sampling point for 2–3 min and mean oxygen saturation values were taken when signals were stable to calculate the depth profiles. For the two-point calibration of the oxygen sensors, Ar-flushed (0% oxygen saturation) and air-purged (100% oxygen saturation) local bottom water was used. High-resolution (1 mm) vertical profiles of oxygen concentrations across the sediment/water interface were accomplished for MUC cores by use of a micromanipulator down to a maximum sediment depth of 5–6 cm. For oxygen measurements in deeper parts of the MUC core as well as for all GCs, holes were drilled through the walls of the core liners in intervals of 1 cm for MUCs and of 5 cm for GCs for the insertion of the microelectrode.

3.3. Pore-water analyses

In the home laboratory at the Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research in Bremerhaven (AWI), NO_3^- was determined using a QuAAtro Continuous Segmented Flow Analyzer (Seal Analytical) with a detection limit of 1.9 μM . Based on duplicate measurements of NO_3^- , the accuracy of the analysis was determined to be < 4.9%. Dissolved manganese (Mn^{2+}) was determined in the acidified pore-water subsamples by inductively coupled plasma optical emission spectrometry (ICP-OES; IRIS Intrepid ICP-OES Spectrometer, Thermo Elemental) with a detection limit of 0.05 μM . Based on the triplicate determination of each sample the reproducibility was < 1.5% for Mn^{2+} .

3.4. Solid-phase analyses

To avoid any interference of the salt matrix in the pore water on the sediment composition, bulk sediment data, total organic carbon and total sulfur contents have been corrected post-analytically according to Kuhn (2013) with the mass percentage of the saline pore water (w') and the mass percentage of H_2O of the wet sediment (w) with the pore water containing 96.5% H_2O (Eq. (1)). With the mass of the salt s [%] (Eq. (2)) the solid-phase composition c' has been calculated using the measured solid-phase composition c (Eq. (3)).

$$w' = w \cdot 100 / 96.5 \quad (1)$$

$$s = 100 \cdot \frac{w' - w}{100 - w} \quad (2)$$

$$c' = c \cdot \frac{100}{100 - s} \quad (3)$$

3.4.1. Total organic carbon content

The content of total organic carbon (TOC) was analyzed using an Eltra CS2000. About 100 mg of freeze-dried, homogenized sediment were weighed into a ceramic cup. Samples were decalcified with 0.5 mL 10% HCl at 250 °C for 2 h before analysis. As the total carbon (TC) consists mostly of TOC in the analyzed samples, total inorganic carbon (TIC) occurs only in negligible amounts. Based on an in-house reference material precision of the analysis was determined to be < 3.7% ($n = 83$).

3.4.2. Radioisotope analyses of ^{231}Pa and ^{230}Th

For the isotope dilution analysis by Inductively Coupled Plasma-Sector Field-Mass Spectrometry (ICP-SF-MS, Element2, Thermo Scientific) freeze-dried and homogenized sediment samples were spiked with about 9 pg ^{229}Th , 0.7 pg ^{233}Pa and 800 pg ^{236}U as internal standards and weighed out in Teflon vials. Total acid digestions were performed in the microwave system MARS Xpress (CEM) according to the procedure described by Kretschmer et al. (2010) and Nöthen and Kasten (2011). Acids were of sub-boiling distilled (HNO_3 , HCl) or suprapur® (HF) quality. About 50 mg of freeze-dried and homogenized bulk sediment were digested in an acid mixture of 65% HNO_3 (3 mL), 30% HCl (2 mL) and 40% HF (0.5 mL) at ~ 230 °C. The digested solutions were fumed off to dryness with the microwave evaporation accessory (CEM XpressVap) and re-dissolved under pressure in 1 M HNO_3 (5 mL) at ~ 200 °C. The residue was filled up to 50 mL with 1 M HNO_3 . After the total acid digestion, 80% of the total digest volume were co-precipitated with $\text{Fe}(\text{OH})_3$. Separation of Pa, Th and U was performed by ion exchange chromatography with the Anion Exchange Resin AG® 1-X8 (Bio-Rad) after the protocols of Anderson and Fleer (1982) and Andersson and Schöberg (2012). Poly-Prep® gravity flow columns filled with AG® 1-X8 resin were conditioned with 9 M HCl (3 * 4 mL) before loading the sample and eluting Th with 9 M HCl (3 * 4 mL), Pa with 9 M HCl/0.14 M HF (4 * 3 mL) and U with 0.1 M HCl (4 * 3 mL), successively. All acids were of sub-boiling distilled or suprapur® quality. The Th, Pa and U eluates were collected and evaporated in Teflon beakers. The Th eluates were purified on a second column with 9 M HCl (400 μL and 6 * 2 mL) for Th, 9 M HCl/0.14 M HF (6 * 2 mL) for Pa and 0.1 M HCl (3 * 4 mL) for U. Separated Th, Pa and U fractions were evaporated, redissolved in HNO_3 and diluted to 1 M HNO_3 for the isotope dilution analysis using ICP-SF-MS with the desolvation system Apex Q (ESI). Data correction for the formation of thorium hydride ($^{232}\text{ThH}^+$), ^{232}Th peak tailing and the instrument mass bias were assessed as described by Kretschmer et al. (2011). Based on the reference material Urem-11 (SARM-31) average accuracy and precision were 0.5% and 1.2% for ^{235}U , 2.4% and 1.2% for ^{230}Th and 0.3% and 2.6% for ^{231}Pa , respectively ($n = 5$).

Sedimentation rates were calculated following the algorithm by Faure (1977), where t is the age[yr] of the sediment and $\frac{^{230}\text{Th}_{ex}}{^{231}\text{Pa}_{ex}}$ is the specific activity ratio at a certain sediment depth x . Here, we used the well constrained value in the bioturbated layer above (Supplementary Fig. 1) as the starting value b , respectively (Eq. (5)).

$$t = \frac{1}{82574.86} \ln \left[\frac{\left(\frac{^{230}\text{Th}_{ex}}{^{231}\text{Pa}_{ex}} \right)_x}{\left(\frac{^{230}\text{Th}_{ex}}{^{231}\text{Pa}_{ex}} \right)_b} \right] \quad (5)$$

Mass accumulation rates [$\text{g cm}^{-2} \text{ kyr}^{-1}$] were determined by the product of sedimentation rate [cm kyr^{-1}] and dry bulk density (DBD) [g cm^{-3}].

3.5. Geochemical model setup and reaction network

A one-dimensional steady-state reaction transport model (e.g. Boudreau, 1997) that couples reactions through a discretized steady-state reaction-transport equation was used to interpret the sedimentary geochemistry at the various CCZ sites (Eq. (6)):

$$0 = \frac{\partial D_{i,j} \vartheta_i (\partial C_{i,j} / \partial z)}{\partial z} - \frac{\partial \omega_i \vartheta_i C_{i,j}}{\partial z} + \alpha_i \vartheta_i (C_{i,j} - C_{0,j}) + \vartheta_i \sum R_{i,j} \quad (6)$$

where z is sediment depth, and i, j represent subscripts depicting depth- and species-dependence, respectively. C is the species concentration (aqueous or solid species, Supplementary material Table 2); D is the diffusive mixing coefficient taking tortuosity (Boudreau, 1997) and bioturbation (Eq. (7)) into account ($D_i = B_i + D_{m,i}$); ϑ is the volume fraction for the aqueous (i.e. the porosity φ) or solid ($1 - \varphi$) phases; ω is

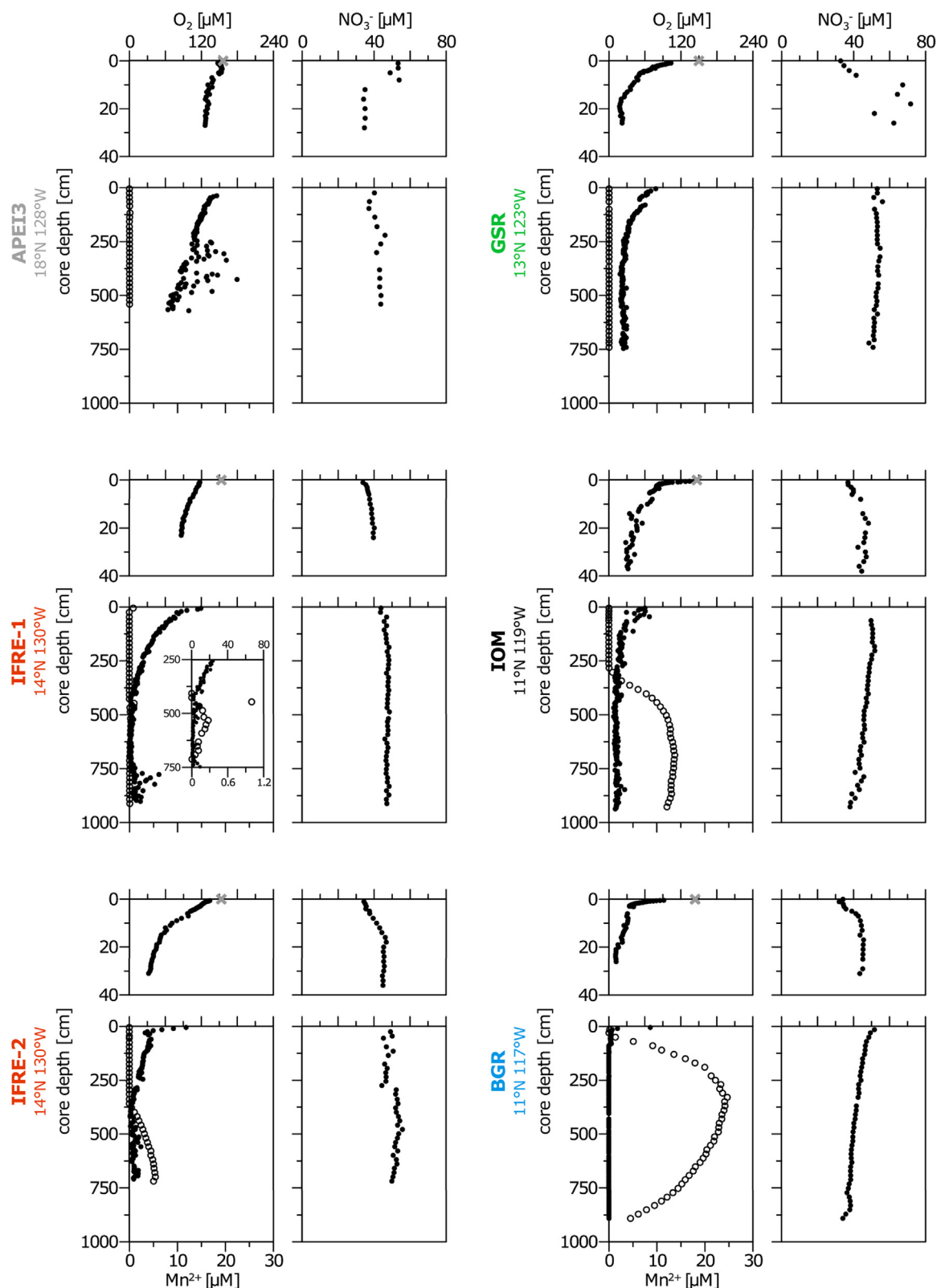
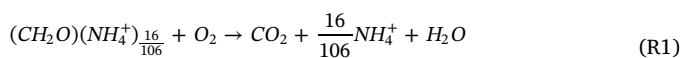


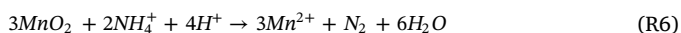
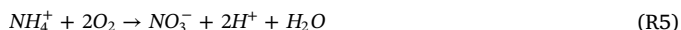
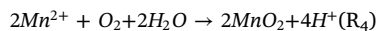
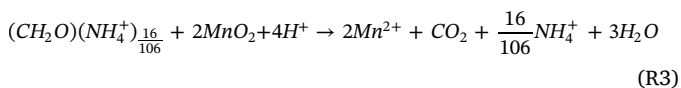
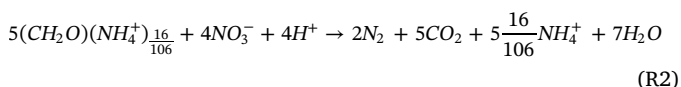
Fig. 2. Pore-water profiles of oxygen, dissolved Mn(II) and nitrate of the APEI3, IFREMER, GSR, IOM and BGR sites. The inset plot for the IFRE-1 site show oxygen (black dots) and dissolved Mn(II) concentrations (open circles) on separate axes (upper axis: oxygen; lower axis: Mn(II)) between 2.5 and 7.5 m sediment depth. Bottom-water oxygen concentrations measured by CTD are indicated (grey crosses).

the velocity of either the aqueous or the solid phase (v or w respectively); α_i is the bioirrigation coefficient (0 for solid species, Eq. (7)); $\sum R_{i,j}$ is the sum of the reactions affecting the given species j .

The reaction-transport model consists of 8 geochemical species and

6 reactions (Supplementary material Table 1):





The chemical reactions during organic carbon degradation were assumed to follow the Redfield ratio stoichiometry with the organic-bound nitrogen to carbon ratio of 16:106 (Redfield, 1934). Biologically induced mixing profiles were assumed to follow a modified logistic function where the break attenuation depths were assumed to be the same for both bioirrigation and bioturbation:

$$B_i = B_0 \exp\left(\frac{z_{\text{mix}} - z}{z_{\text{att}}}\right) / \left(1 + \exp\left(\frac{z_{\text{mix}} - z}{z_{\text{att}}}\right)\right) \quad (7)$$

$$\alpha_i = \alpha_0 \exp\left(\frac{z_{\text{mix}} - z}{z_{\text{att}}}\right) / \left(1 + \exp\left(\frac{z_{\text{mix}} - z}{z_{\text{att}}}\right)\right) \quad (8)$$

where α_0 and B_0 are constants representing the maximum bioirrigation and bioturbation coefficients at sediment-water interface, z_{mix} is the depth to where α_0 and B_0 become half their value and z_{att} is the attenuation of the biogenically induced mixing with depth.

Porosity data were fit for each station and showed an exponential decrease with depth typical of compacting sediment (not shown) and were fit with the following equation:

$$\varphi_i = \varphi_\infty (\varphi_0 - \varphi_\infty) \exp(-\beta z) \quad (9)$$

where φ_∞ is the porosity at compaction, φ_0 is the porosity at the sediment water interface, and β is the depth-attenuation coefficient. Sediment thicknesses for the diagenetic model were extracted from NCEI's global ocean sediment thickness grid (Whittaker et al., 2013). For the BGR site, sediment thicknesses were inferred from nearby core locations with similar pore-water Mn profiles (SO240-9KL, SO240-96SL; Kuhn et al., 2017). The model was coded in R (version 3.2.4) using the ReacTran package (Soetaert and Meysman, 2012) to solve Eq. (6) and the marelac package (Soetaert et al., 2010) to solve the molecular diffusion coefficients for the modeled species ($D_{m,i}$). The advective velocities of solid and pore water phases were solved using the compact grid function within the ReacTran package, which takes sedimentary compaction into account.

4. Results

4.1. Core description

All study sites show light brown clay-dominated siliceous ooze with variable surface nodule coverage (Table 1). The sediments in the APEI3 site are dominated by dense and dry, dark brown sediment with a comparably low degree of lithological variation. All other sites are characterized by light brown sediments with irregular dark patches and layers throughout.

4.2. Pore water

Bottom-water oxygen concentrations measured with the CTD (SBE 43 oxygen self-regenerative WetLabs Clark-sensor) are 156 μM in the APEI3 area, 153 μM in the IFREMER area, 150 μM in the GSR area, 147 μM in the IOM area and 144 μM in the BGR area (Fig. 2). In the surface sediments, oxygen concentrations usually decrease rapidly with depth in the upper 30 cm (Fig. 2). However, in contrast to the other

sites, the surface sediments of the APEI3 and IFRE-1 sites show only a slight decrease in oxygen. Oxygen concentrations below detection limit are reached at various sediment depths while the sediments at the APEI3 and the GSR sites remain oxic throughout. The oxygen penetration depth (OPD) is 1 m at the BGR site, 3 m at the IOM site, 4.5 m at the IFRE-1 site and 3.8 m at the IFRE-2 site.

Pore-water Mn^{2+} occurs below the OPD within a wide concentration range of 0.2–25 μM (Fig. 2). At the IFRE-1 site, Mn^{2+} concentrations < 1 μM are between 4.5 and 7 m while the IFRE-2 site shows a concave-up profile with higher concentrations of up to 5.5 μM . At the IOM site, Mn^{2+} increases downward to 13 μM and slightly decreases below 8 m. Overall, the highest Mn^{2+} concentrations were measured at the BGR site with up to 25 μM at 3.3 m and a subsequent decrease in concentrations with depth.

NO_3^- concentrations generally increase with depth from bottom water concentrations of 35 μM at the sediment surface to 45 μM in the upper 30 cm (Fig. 2). The APEI3 and GSR sites show NO_3^- concentrations in the surface sediments of up to 53 μM and 70 μM , respectively. A decrease of NO_3^- with depth below 30 cm is detected at the BGR, IOM and IFRE-2 sites to about 35 μM at the bottom of the cores. At the APEI3, IFRE-1 and GSR sites, NO_3^- concentrations remain mostly constant throughout the deep sediment core.

4.3. Solid phase

4.3.1. Total organic carbon

The TOC contents generally decrease with depth in the upper 30 cm (Fig. 3). The lowest surface sediment TOC contents of 0.2 wt% are found in the APEI3. Both, the IFRE-1 and IFRE-2 sites, show 0.3–0.4 wt% of TOC while the GSR site and the IOM site have 0.5 wt% of TOC. The highest contents with 0.6 wt% are found at the BGR site. Below 30 cm, the TOC remains < 0.2 wt% (Fig. 3). A drop to TOC contents of < 0.1 wt% is found at the GSR site below 5 m sediment depth.

4.4. Sedimentation rates and bioturbation depth

Sedimentation rates range between 0.2 and 1.15 cm kyr^{-1} (Fig. 4). The rates at the APEI3 and the GSR sites are at least threefold lower than at the IFRE-1 and the IOM sites. Bioturbation is usually limited to the upper 7 cm whereas the bioturbated layer at the IOM site reaches down to 13 cm (Supplementary Fig. 1).

5. Discussion

5.1. POC flux to the seafloor and sedimentation rates

Pelagic deep-sea sediments receive little organic matter (OM) due to generally low surface water productivity and great water depth (Heath et al., 1977; Müller and Suess, 1979; Honjo, 1980). For the prediction of carbon export production at any water depth, Suess (1980) developed an empirical algorithm from sediment trap measurements and primary production (PP) rates in the respective surface waters. Several studies have applied modified algorithms after Suess (1980) (e.g., Dymond et al., 1997; Tyrell, 1999) and Martin et al. (1987) (e.g., Emerson et al., 1997; Fischer et al., 2000) for predicting the vertical POC flux. However, these commonly applied relationships between POC flux and water depth generally overestimate the flux of OM to depth (Buesseler et al., 2007; Buesseler and Boyd, 2009; Henson et al., 2011; Arndt et al., 2013 and references therein). Lutz et al. (2002) developed regional algorithms including region-specific sinking and remineralization rates, i.e. labile and refractory POC fractions in the water column. Using this empirical parameterization of the POC flux to depth, Lutz et al. (2007) combined a time series of remotely sensed net PP in the surface waters, sea surface temperature and sediment trap POC flux data to construct models with global predictions of POC fluxes to the seafloor. According to the model by Lutz et al. (2007), about 1.7 $\text{mg C}_{\text{org}} \text{m}^{-2} \text{d}^{-1}$ is delivered

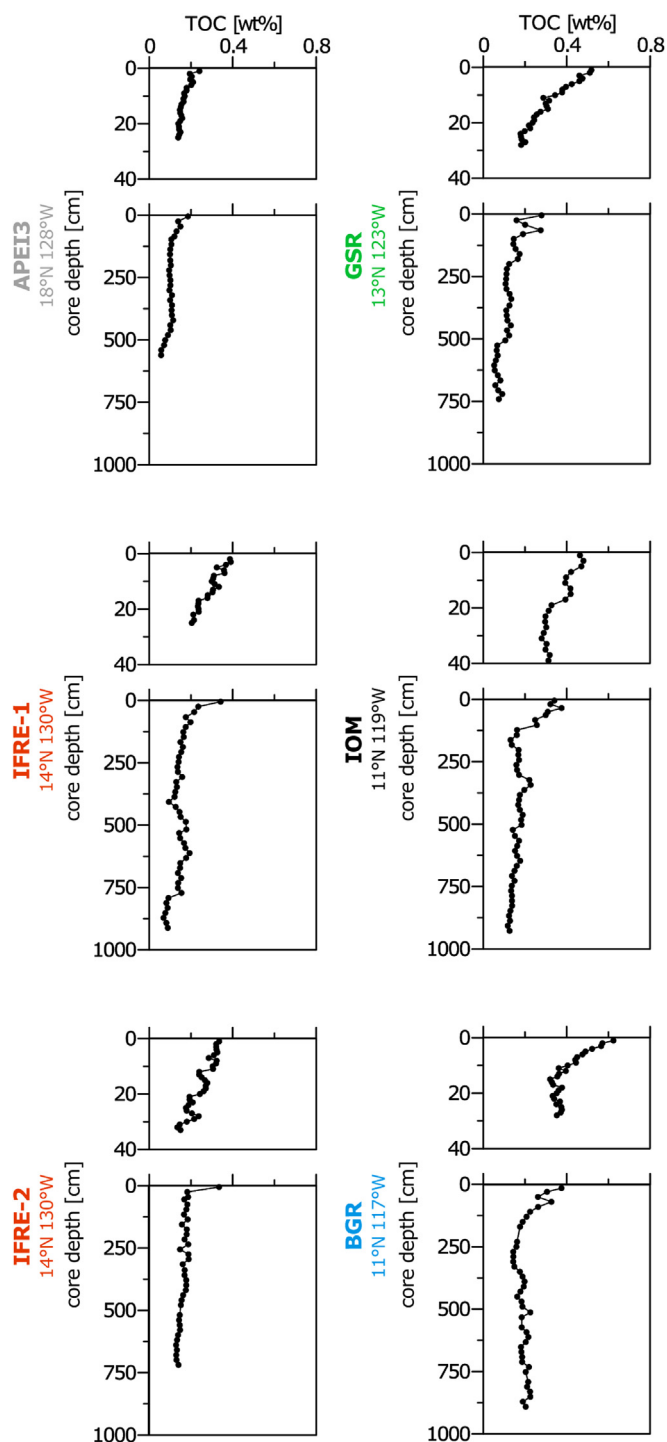


Fig. 3. Solid-phase profiles of TOC of the APEI3, IFREMER, GSR, IOM and BGR sites.

to the seafloor at the BGR and IOM sites, $1.5 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$ reaches the sediments of the GSR and IFREMER sites and $1.3 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$ settles to the seafloor in the APEI3 area (Fig. 1). These fluxes may be biased due to (1) significant uncertainties in the NPP estimates based on satellite data and (2) several potential errors in the trapping efficiency of sediment traps (Lutz et al., 2007 and references therein). Furthermore, with water depths of 4–5 km, the export production may be laterally drifted during settling, especially by strong bottom currents in the proximity of seamounts (e.g., Mewes et al., 2014). We implemented the POC fluxes given by Lutz et al. (2007), which can be regarded as rough estimates, into our reaction transport model and adjusted these values

in order to fit the measured profiles of TOC and the oxidants. Our diagenetic model reproduces the Lutz et al. (2007) fluxes within 20% (Table 2). Only a slight discrepancy occurs between the POC fluxes derived from both models for the APEI3 and BGR sites. Compared to the other studied contract areas, the water column in the APEI3 area is characterized by a less pronounced OMZ (Martínez Arbizu and Haeckel, 2015). Thus, due to the significantly longer oxygen exposure time of settling POC, which is the key parameter determining the degradation efficiency of POC (e.g. Hartnett et al., 1998; Banse, 1990; Zonneveld et al., 2010; Cavan et al., 2017), the degradation in the water column may be enhanced in this area. The regional differences in the OMZ thickness may limit the empirical algorithm used by Lutz et al. (2007) for calculating the POC flux to the seafloor. Thus, POC fluxes in the APEI3 area are most likely overestimated. Conversely, the POC flux in the BGR area has likely been underestimated as the water column is characterized by an extensive OMZ (Martínez Arbizu and Haeckel, 2015) aiding the POC preservation in the water column and triggering a higher POC flux to the sediment (Table 2). This is consistent with our 20% lower fluxes at the APEI3 site and 20% higher POC flux at the BGR site in comparison with the Lutz et al. (2007) fluxes.

Although considerably higher estimated POC fluxes between 1.5 and $11.5 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$ have been reported from nearby stations (Murray and Kuivila, 1990; Mogollón et al., 2016), the TOC contents at the sediment surface at all sites are mostly in agreement with published values from the CCZ ranging from 0.1 to 0.6 wt% (Müller, 1977; Müller and Mangini, 1980; Jahnke et al., 1982; Murray and Kuivila, 1990; Mewes et al., 2014, 2016). Mewes et al. (2014) have presented geochemical data for sediments from the BGR contract area “East” with two sites close ($< 5 \text{ km}$) to a large seamount between the BGR and IOM area (A1-1-MN, A1-2-NN) and two sites far away from seamounts (A5-1-BN, A5-2-SN). While the surface sediment TOC contents are similar at all stations, a comparably rapid decrease in TOC contents with depth occurs at the sites adjacent to the seamount with relatively high POC fluxes of $10 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$ (Fig. 6; Mogollón et al., 2016). Our BGR site shows up to 20% higher TOC contents in the upper 30 cm than the sites studied by Mewes et al. (2014) which also decrease rapidly with sediment depth (Fig. 6). Khrpounoff et al. (2006) reported 25% higher TOC contents in the upper 30 cm at a site in the IFREMER area (Fig. 6). Variations in local estimated POC fluxes, TOC contents and sedimentation rates are potentially a result of strong local heterogeneities of bottom water currents which can be intensified, attenuated and deflected in the vicinity of seamounts and ridges (Hogg, 1973; Gould et al., 1981; Mohn and Beckmann, 2002; Xu and Lavelle, 2017; Juan et al., 2018) and cause kilometer-scale differences in the supply and composition of sediments (Turnewitsch et al., 2004, 2015; Mewes et al., 2014). The relatively high sedimentation rates at the IOM site of $1.15 \text{ cm} \cdot \text{kyr}^{-1}$ may be associated with sediment focusing. Furthermore, sediments of different composition may be delivered from the slopes of nearby seamounts and ridges by gravity-induced processes (e.g., Stanley and Taylor, 1977; Jeong et al., 1994).

5.2. Organic matter degradation

Müller et al. (1988) have suggested that aerobic respiration is the dominant biogeochemical process degrading sedimentary OM in the CCZ and previous modeling studies for the BGR contract area “East” (Table 2; Fig. 5; Mewes et al., 2016; Mogollón et al., 2016) have supported this assumption. Our model simulations are in line with these findings and show that aerobic respiration consumes more than 90% of the OM delivered to the seafloor while denitrification and Mn(IV) reduction together consume less than 1%. These numbers agree well with rates derived from the previous modeling studies for the BGR contract area “East” (Table 2; Fig. 5; Mogollón et al., 2016; Mewes et al., 2016). The zone in which manganese and nitrate reduction co-occur will hereafter be referred to as “suboxic zone” in which neither oxygen nor hydrogen sulfide are present (Bernier, 1981). Most of the ammonia

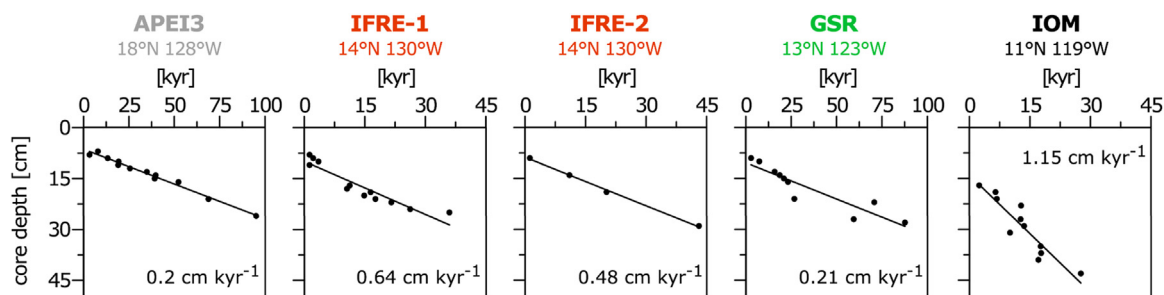


Fig. 4. $^{230}\text{Th}/^{231}\text{Pa}$ -derived sedimentation rates for the uppermost 50 cm of the sediments of the APEI3, IFREMER, GSR and IOM sites.

Table 2

Fluxes of POC and oxygen into the sediment and depth-integrated rates of the predominant biogeochemical processes derived from the diagenetic model.

Biogeochemical process	Unit	BGR	IOM	GSR	IFRE-1	IFRE-2	APEI3
POC flux	[mg C m ⁻² d ⁻¹]	1.99	1.54	1.51	1.47	1.5	1.07
O ₂ flux top	[mg O ₂ m ⁻² d ⁻¹]	5.18	4.24	3.61	4.3	4.37	2.58
O ₂ reduction	[mg C m ⁻² d ⁻¹]	1.92	1.44	1.38	1.3	1.45	0.93
NO ₃ ⁻ reduction	[mg C m ⁻² d ⁻¹]	1.5E-03	1.7E-03	3.3E-02	5.5E-03	5.0E-03	8.7E-04
Mn(IV) reduction	[mg C m ⁻² d ⁻¹]	1.5E-03	6.1E-04	1.6E-02	5.7E-04	4.6E-03	1.4E-03
Nitrification	[mg N m ⁻² d ⁻¹]	0.01	0.13	0.14	0.10	0.14	0.03
Mn ²⁺ oxidation	[mg O ₂ m ⁻² d ⁻¹]	0.062	0.026	0.06	0.025	0.029	0.016
Mn-annamox	[mg Mn m ⁻² d ⁻¹]	0.14	0.005	0	0	0	0

liberated from OM degradation becomes nitrified in the oxic zone so that the nitrate concentrations increase with depth in the upper 30 cm of sediment (Fig. 2; Fig. 5). Additionally to nitrification, the breakup of nitrate-rich organic compounds may contribute to the relatively high NO₃⁻ concentrations in the surface sediments at the APEI3 and GSR sites (e.g., Jørgensen and Gallardo, 1999). The bioturbated layer includes the upper 6–13 cm of the oxic sediment which is in good agreement with the average bioturbation depth of 10 cm (Boudreau, 1994). The determined low sedimentation rates of 0.2–1.15 cm kyr⁻¹ in combination with low POC fluxes to the seafloor (< 2 mg C_{org} m⁻² d⁻¹) lead to low carbon burial rates of less than 0.15 mg C_{org} m⁻² d⁻¹ (based on depth-integrated rates, cf. Table 2), which are consistent with other organic carbon burial estimates for the CCZ (e.g., Jahnke, 1996). The highly refractory TOC fraction of < 0.2 wt% below 20 cm is insufficient to reduce nitrate (Table 2; Fig. 5; Supplementary Table 2; Mogollón et al., 2016). These observations are in agreement with Müller and Mangini (1980) and Müller et al. (1988) who state that with sedimentation rates of 0.2–0.4 cm kyr⁻¹ OM is almost completely remineralized within the uppermost meter of the sediments. Mewes et al. (2014) determined carbon respiration rates for sediments in the BGR contract area “East” that showed twofold higher respiration rates in the upper oxic sediments compared to the depth-integrated rates of aerobic respiration derived from our diagenetic model (Table 2). This offset may be caused by the alteration of the microbial communities during sediment retrieval, i.e. decompression (e.g., Park and Clark, 2002), and the different approaches of respiration rate determination. Moreover, the discrepancy in the respiration rates may reflect the lateral heterogeneity in the supply of TOC related to the interaction of currents with rough seafloor topography, in particular seamounts. Twofold higher rates of aerobic respiration at the BGR site compared to the APEI3 site are a result of twofold higher POC fluxes to the seafloor at the BGR site than at the APEI3 site in the proximity of the carbon-starved North Pacific gyre. Consequently, the APEI3 site does not represent the (bio) geochemical conditions of the sites in the European contract areas investigated in the framework of this study. This observation is in agreement with Vanreusel et al. (2016) who explained the significantly lower faunal densities in the APEI3 with primary productivity being much lower compared to in the contract areas.

5.3. Redox zonation and oxygen fluxes

The *ex situ* sediment oxygen concentrations are in good agreement with other oxygen data for the equatorial Pacific Ocean (Murray and Grundmanis, 1980; Jahnke et al., 1982; Berelson et al., 1990; Hammond et al., 1996; Khripounoff et al., 2006; Mewes et al., 2014, 2016; Rühlemann et al., 2011). In accordance with oxygen measurements by Mewes et al. (2014), bottom water oxygen measurements of the overlying water of the MUC cores using amperometric Clark-type oxygen sensors are systematically 10–20% higher than bottom-water concentrations determined with the CTD 20–30 m above the seafloor (Martínez Arbizu and Haeckel, 2015). This offset is most likely a result of sampling artifacts that occurred during retrieval of the MUC cores or of atmospheric oxygen diffusion into the bottom water of the surface sediments during *ex situ* measurements, which biases the oxygen concentrations down to about 0.4 cm sediment depth. The threshold of 0.4 cm has been determined in this study by triplicate profiling across the water-sediment interface resulting in systematically higher oxygen concentrations of each consecutively measured profile. Therefore, we excluded oxygen concentrations in the uppermost 0.4 cm from the calculation of oxygen gradients and fluxes and used the CTD oxygen concentrations as bottom water values (Table 2; Table 3). The oxygen data obtained for the IOM site indicate that oxygen is not completely depleted throughout the sediment core (Fig. 2). However, the detection of dissolved Mn²⁺ in pore water below 3 m sediment depth suggests that oxygen is absent in this interval. We attribute this discrepancy to some calibration problems of one of the sensors at the beginning of the cruise which affected the oxygen measurements at the BGR and IOM sites. Additionally, the discrepancy between the GSR oxygen concentrations of the MUC and the GC is attributed to small-scale variations or to miscalibration of the oxygen sensor. Even though the oxygen data shows sampling artifacts and miscalibration, which partly bias the absolute oxygen concentrations, it can be used in combination with the pore-water Mn²⁺ data to delineate the sedimentary redox zonation. As the oxygen data at the bottom water-sediment interface is particularly affected, oxygen fluxes into the sediment are given with an error of < 5%.

The OPD inversely correlates with the POC flux ($r^2 = 0.85$; Fig. 7). The OPD of 1 m at the BGR site is shallower than the OPD reported for other sites studied in this region which range between 1.8 and 3 m

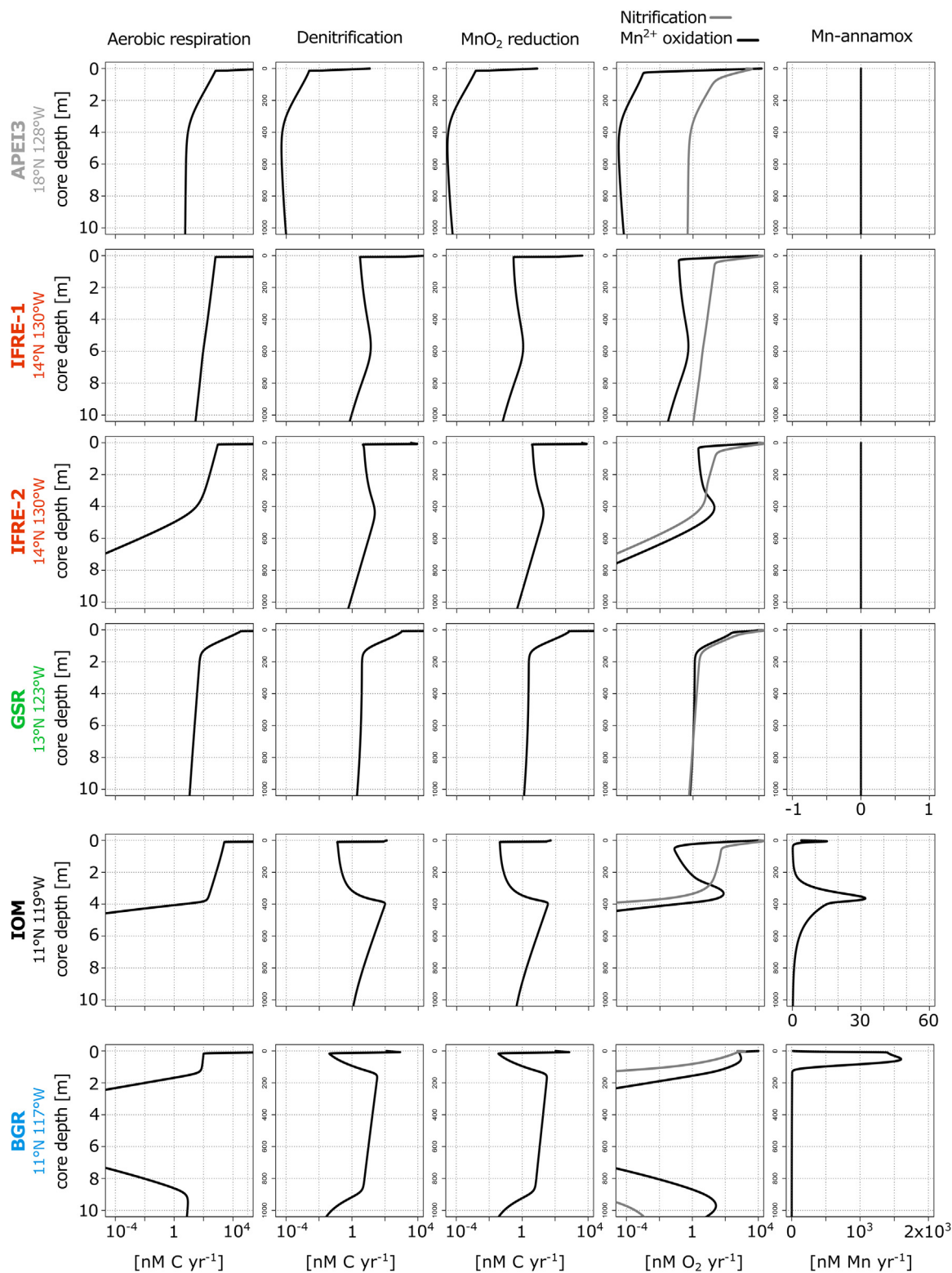


Fig. 5. Model results for all sites including the biogeochemical processes of aerobic respiration, Mn(IV) reduction, denitrification, Mn^{2+} oxidation, nitrification and Mn-annamox (R_1 - R_6).

depth (Rühlemann et al., 2011; Mewes et al., 2014). Such small-scale variation is reflected in the oxygen gradients with 2.6 – $5.6 \mu\text{M O}_2 \text{ mm}^{-1}$ in the sediments in the BGR area (Table 3). OPDs are generally deeper at sites with medium-sized or big (3–10 cm) nodules at the sediment surface than at sites with small-size (1–8 cm) or without nodules at the surface (Mewes et al., 2014). The two IFREMER sites (IFRE-1 and IFRE-

2) are ~ 6 km apart with medium-sized surface nodules at the IFRE-1 site and no nodules at the IFRE-2 site (Table 1) and in accordance with the findings of Mewes et al. (2014), the OPD at the IFRE-1 site is located 70 cm deeper than at the IFRE-2 site (Fig. 2). However, the depth-integrated rates of Mn(IV) and nitrate reduction are not significantly higher at the IFRE-2 site compared to the IFRE-1 site (Table 2; Fig. 5)

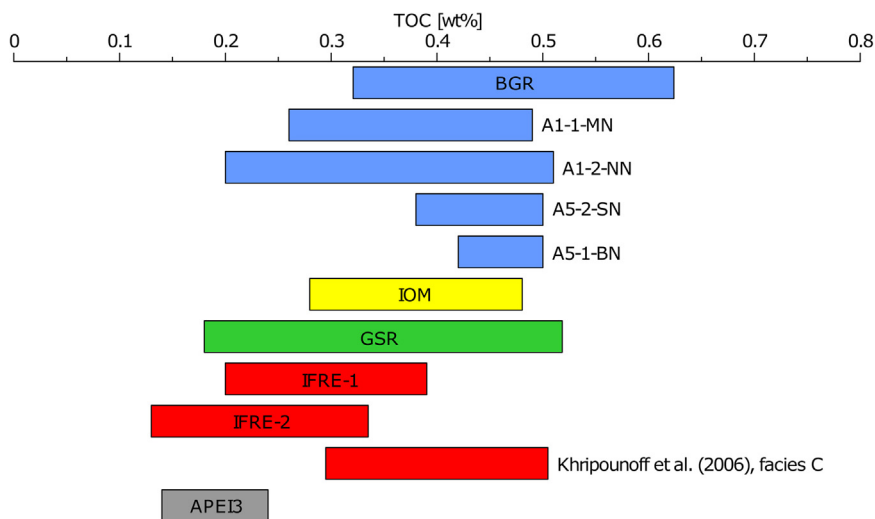


Fig. 6. Comparison of the TOC contents of surface sediments (upper 30 cm) within the different contract areas CCZ. The data for the BGR sites A1-1-MN, A1-2-NN, A5-2-SN, A5-1-BN from cruise SO205 are taken from Mewes et al. (2014). The range of TOC contents comprises the lowest to highest measured contents throughout MUC cores taken at the individual sites. The highest TOC content at each site was determined at the sediment surface, i.e. in the uppermost centimeter of the sediment.

Table 3

Oxygen gradients calculated from oxygen profiles for the uppermost 0.4–1 cm of sediment. Negative gradients indicate the oxygen flux into the sediment. The data from SO205 were taken from Mewes et al. (2014). Note that the oxygen gradient at the GSR site is biased due to miscalibration of the sensor.

Station	Oxygen gradient [$\mu\text{M mm}^{-1}$]
SO205–05MUC	– 3.5
SO205–06MUC	– 2.6
SO205–46MUC	– 5.6
SO205–48MUC	– 9
BGR	– 5
IOM	– 7
GSR	– 8.3
IFRE-1	– 3.5
IFRE-2	– 1.7
APEI3	– 0.2

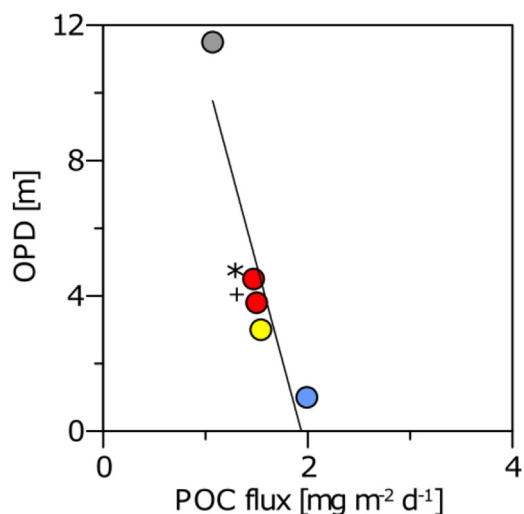


Fig. 7. Correlation between the oxygen penetration depth (OPD) and the POC fluxes used as boundary conditions for the diagenetic model. Based on interpolation of the oxygen profile at the APEI3 site, oxygen may be consumed at 11.5 m. The linear regression through the BGR (blue), IOM (yellow), *IFRE-1 (red), +IFRE-2 (red) and APEI3 (grey) sites shows a coefficient of determination of $r^2 = 0.85$.

which disagrees with the results presented by Mewes et al. (2014). This discrepancy may be connected to almost twofold higher sedimentation rates proposed by the authors for the nodule-free sites compared to the sites with medium-sized/big nodules while in the IFREMER area, sedimentation rate at the IFRE-1 site exceeds the rate at the IFRE-2 site. In accordance with the low respiration rates at the APEI3 site, the oxygen gradient over the upper 0.4–1 cm of sediment is low at $0.2 \mu\text{M O}_2 \text{ mm}^{-1}$ (Table 3). At depth, oxygen concentrations are scattered at the APEI3 site. We interpret this as an analytical artifact because the sediments at the APEI3 site were very dry and consolidated and have therefore very likely impacted the sensor measurements.

In the suboxic zone, Mn^{2+} concentrations between 0.2 and $25 \mu\text{M}$ agree well with reported values in the CCZ (Emerson et al., 1980; Jahnke et al., 1982; Mewes et al., 2014). Mn^{2+} is mainly mobilized by the dissimilatory reduction of Mn(IV) phases below the OPD (Figs. 2 and 5). As proposed by Mogollón et al. (2016) for adjacent sites, the observed concomitant decrease of nitrate in the suboxic zone of the sediment core from the BGR area suggests that the biogeochemical cycles of manganese and nitrogen are coupled. Excess ammonium produced by OC degradation in the oxic zone may escape aerobic nitrification and diffuse downwards into the suboxic zone where it can act as electron donor during manganese oxide reduction (Mn-annamox, Luther et al., 1997). Mn-annamox may induce the liberation of Mn^{2+} into the pore water of the sediments of the BGR and IOM sites with reaction rates similar to the rates published by Mogollón et al. (2016) (Table 2; Fig. 5). The nitrate concentrations determined within the framework of this study are in correspondence with previously published nitrate pore-water data from the CCZ ranging from 30 to $70 \mu\text{M}$ (Jahnke et al., 1982; Jeong et al., 1994; Mewes et al., 2014, 2016; Mogollón et al., 2016). Nitrate reduction is very weak at these sites due to the low and refractory OM content (Table 2). Iron and manganese oxidation coupled to nitrate reduction is probably also not feasible due to the low Mn^{2+} concentrations and Fe^{2+} levels below detection limit.

5.3.1. Deep Mn^{2+} oxidation

Decreasing-with-depth profiles of Mn^{2+} at the IFRE-1, IOM and BGR sites indicate that Mn^{2+} is consumed by either precipitation of an authigenic carbonate mineral (e.g., Gingele and Kasten, 1994) or by oxidation at depth (Fig. 2). Due to the fact that alkalinity is low in these sediments (data not shown), and thus, precipitation of a Mn carbonate phase is unlikely, we suggest that at depth, Mn^{2+} is most likely oxidized by oxygen diffusing from the underlying basaltic basement. Such upward diffusive supply of oxygen from oxic seawater circulating in the oceanic crust was first shown for the German contract area “East” by Mewes et al. (2016) and has recently been documented to be

widespread in the area by Kuhn et al. (2017). At the IFRE-1 site, Mn^{2+} is consumed at 7 m depth. Based on interpolation of the Mn^{2+} profiles, Mn^{2+} is reoxidized at ~ 9.25 m depth at the BGR site while it is consumed at ~ 20 m depth at the IOM site. The low-temperature circulation of seawater through the basaltic crust (e.g., Fisher and Wheat, 2010; Ziebis et al., 2012) underlying the sediments of the CCZ had so far only been shown for the BGR area “East” on the base of increasing-with-depth oxygen and decreasing-with-depth Mn^{2+} profiles (Mewes et al., 2016; Kuhn et al., 2017). Numerous seamounts and faults in the eastern CCZ have been shown to facilitate the recharge and subsequent discharge of oxic seawater into and from oceanic crust and diffusion of oxygen into the overlying sediments. The BGR, IOM, and IFRE-1 sites are located about 5 km, 3 km and 1 km, respectively from the adjacent ridge flanks (Fig. 1). At these sites, the sediment drape thicknesses range between 20 and 83 m (Supplementary Table 2). In comparison, the sites studied by Kuhn et al. (2017) in the BGR area had an average sediment thickness of 48 m. The Mn^{2+} profile at the IFRE-2 site does not indicate Mn^{2+} oxidation at depth while the sediments at the APEI3 and GSR sites are oxic throughout. The extended oxic zones at the APEI3 and GSR sites probably result from low organic carbon supply and respiration (Table 2), which is insufficient to fully consume the oxygen diffusing into the sediment from both the overlying seawater and the underlying basaltic crustal fluids. However, the retrieved core lengths of up to 7.5 m do not allow the identification of deep oxidative processes at these locations. Based on our pore-water data, widespread deep Mn^{2+} oxidation occurs at three sites of various contract areas within the CCZ.

6. Conclusion

We studied six abyssal sites in the Clarion-Clipperton Zone of five European contract areas for the exploration of polymetallic nodules and one site in an Area of Particular Environmental Interest (APEI3) over a distance of about 1300 km. The sites differ in POC fluxes to the seafloor ($1\text{--}2 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$) and sedimentation rates ($0.2\text{--}1.15 \text{ cm kyr}^{-1}$) while the bioturbation depth is mostly limited to the upper 7 cm of the sediment. Solid-phase contents, pore-water profiles and the applied transport reaction model demonstrate significant inter-areal differences in sedimentation rates, the extension of oxic and suboxic zones and rates of organic matter remineralization. We show that the observed variability in redox zonation at the study sites is determined by differences in (1) surface water productivity and associated POC flux to the seafloor, (2) sediment accumulation rate and (3) the oxidation of pore-water Mn^{2+} at depth. Diagenetic modeling indicates that due to the low sedimentation rates, the labile fractions of organic carbon are restricted to the upper 20 cm of the sediment where OC degradation is dominated (90%) by aerobic respiration. Mn(IV) reduction and denitrification each consume less than 1% of the refractory organic matter. The suboxic zone is characterized by a wide range of Mn^{2+} concentrations where the sites with the highest POC fluxes indicate further production of Mn^{2+} by Mn-annamox. Mn^{2+} -mediated denitrification is absent at all sites due to insufficient Mn^{2+} concentrations. Downward decreasing Mn^{2+} concentrations at three sites indicate the widespread oxidation of Mn^{2+} at depth throughout the CCZ. Due to very low POC fluxes of $1 \text{ mg C}_{\text{org}} \text{ m}^{-2} \text{ d}^{-1}$ to the seafloor in the APEI3 area, respiration rates at this site are about twofold lower than in the investigated European contract areas. Consequently, we infer that the preservation area APEI3 does not represent the depositional conditions and biogeochemical processes that are dominating in the investigated European contract areas. The POC fluxes to the seafloor and the sedimentation rates generally correlate with surface water productivity, however, within a given contract area, small-scale spatial (intra-areal) variations in geochemical conditions and biogeochemical processes are caused by (1) various extents of POC degradation processes in the water column and (2) small-scale interactions of bottom water currents and topography affecting the sedimentation pattern.

This study represents the first biogeochemical baseline study that advances our knowledge about regional variations of natural depositional and geochemical conditions as well as biogeochemical processes in the sediments of the vast deep-sea area of the CCZ. Our findings may deliver important baseline data to be used for the assessment of the impact of potential deep-sea mining activities. They will also serve as an input for the further development of the Environmental Management Plan by the International Seabed Authority (ISA).

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.dsr.2018.08.006.

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