

Impact of deep-sea polymetallic nodule mining on benthic microbial community and mediated biogeochemical functions

MASSIMILIANO MOLARI¹, Tobias R. Vonnahme^{1*}, FELIX JANSSEN¹, FRANK WENZHÖFER¹, MATTHIAS HAECKEL², JÜRGEN TISCHACK⁴, ANTJE BOETIUS¹



¹ Joint Research Group on Deep-Sea Ecology & Technology

MPI Bremen | AWI Bremerhaven, DE

² GEOMAR | Kiel, DE

³ MARUM | University of Bremen, DE

* now at UiT Arctic University of Norway | Tromsø, NO



Max Planck Institute
for Marine Microbiology



ALFRED-WEGENER-INSTITUT
HELMHOLTZ-ZENTRUM FÜR POLAR-
UND MEERESFORSCHUNG



GEOMAR
Helmholtz Centre for Ocean Research Kiel



Zentrum für Marine
Umweltwissenschaften

Where to carry out mining in the deep-sea and for what?

Massive sulfide deposits



Cobalt-rich crust

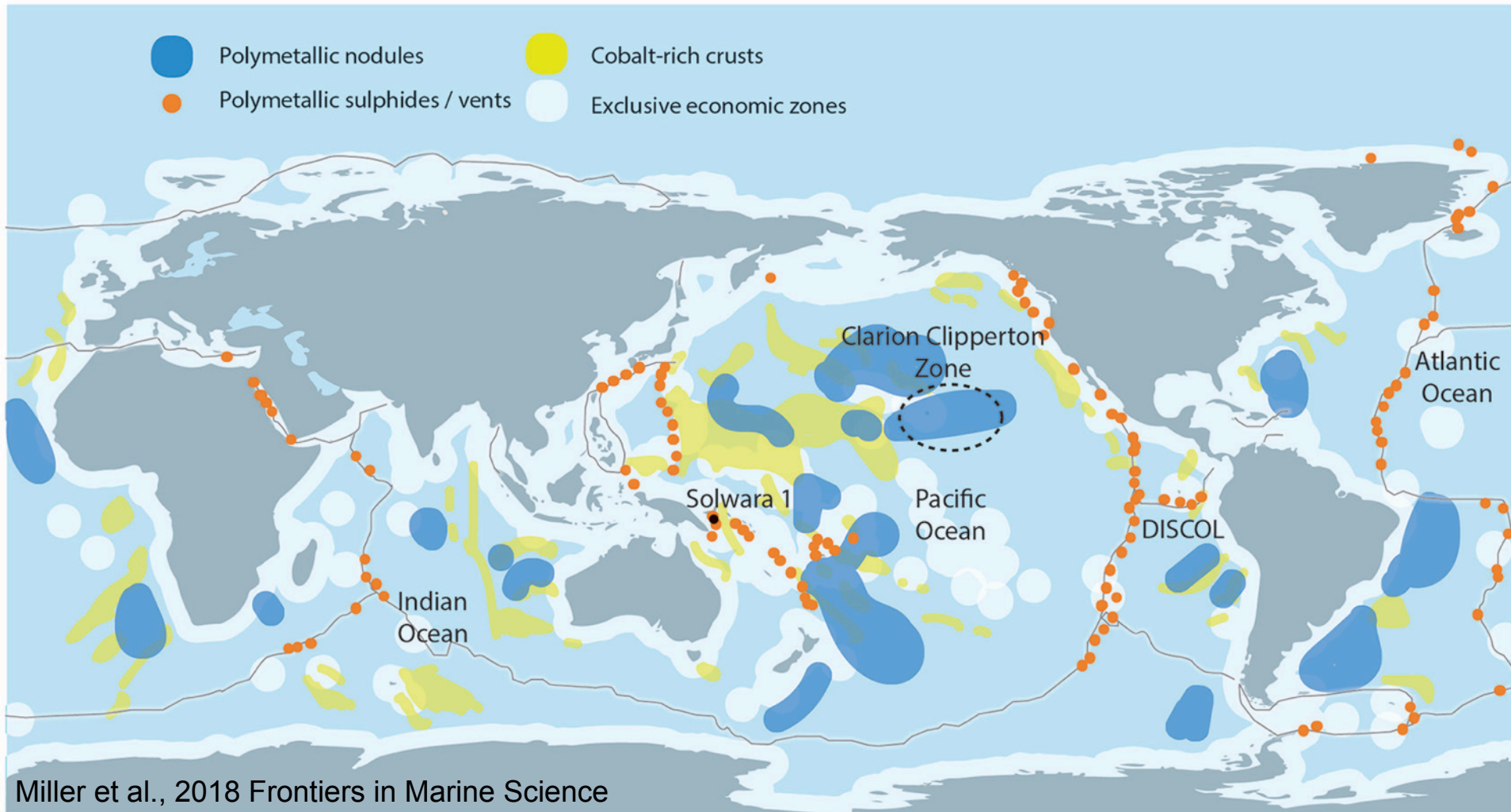


Manganese nodule fields



Metal resources include: arsenic, copper, cobalt, nickel, lithium, platinum, tellurium, zinc, lead, barium, gold, silver and rare earth elements

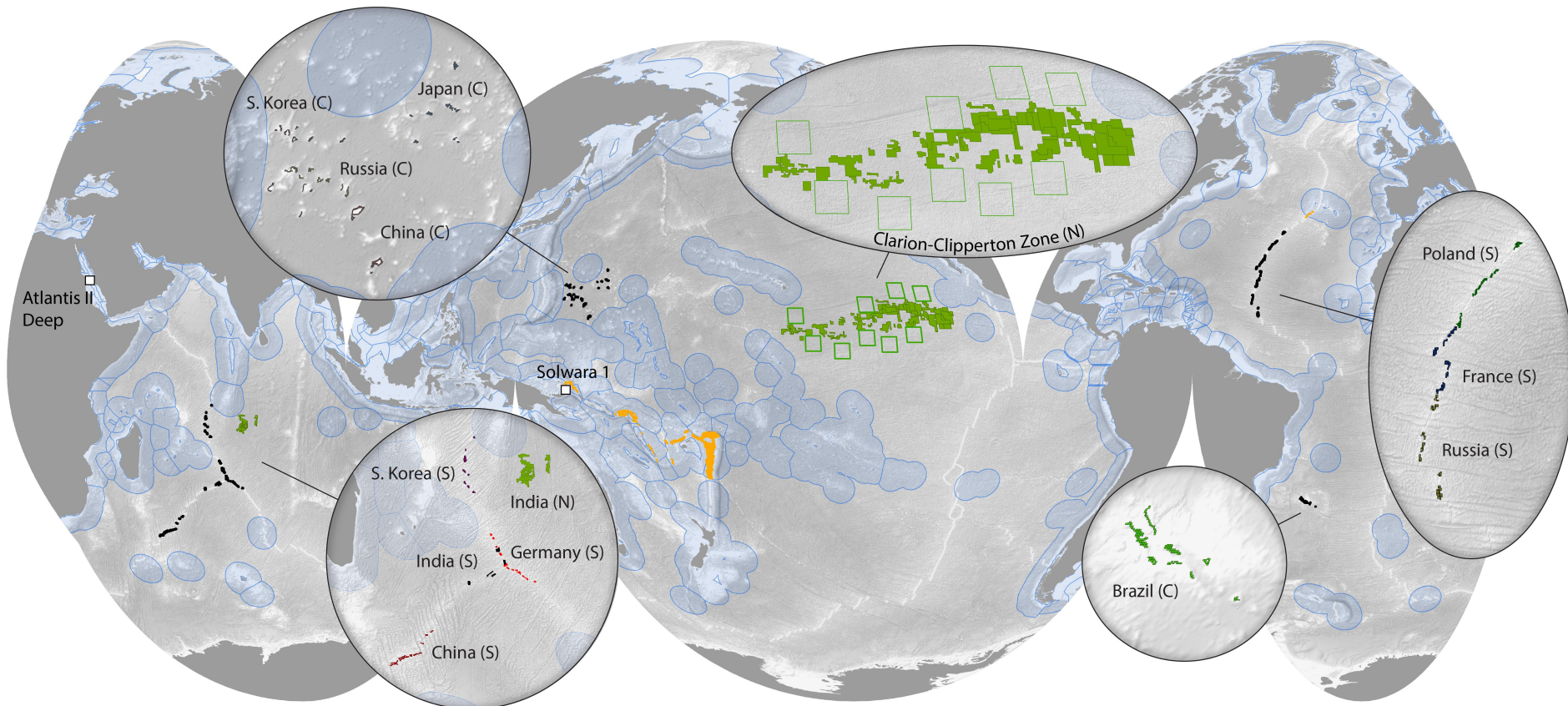
Where to carry out mining in the deep-sea and for what?



Metal resources include: arsenic, copper, cobalt, nickel, lithium, platinum, tellurium, zinc, lead, barium, gold, silver and rare earth elements

Is deep-sea mining ready to start?

International Seabed Authority (ISA) has approved **28 exploration contracts** in the Pacific, Indian, and Atlantic oceans – covering more than **85,000 km² of deep-sea area**



Map: Sven Petersen & Anna Krätschel, GEOMAR

Is deep-sea mining ready to start?

So far not a single request for an exploitation license was handed in, but ISA is working on the regulations and the contractors on the technical preparations:

In 2017 Japan completed the first successful test for zinc extraction from the deep seabed

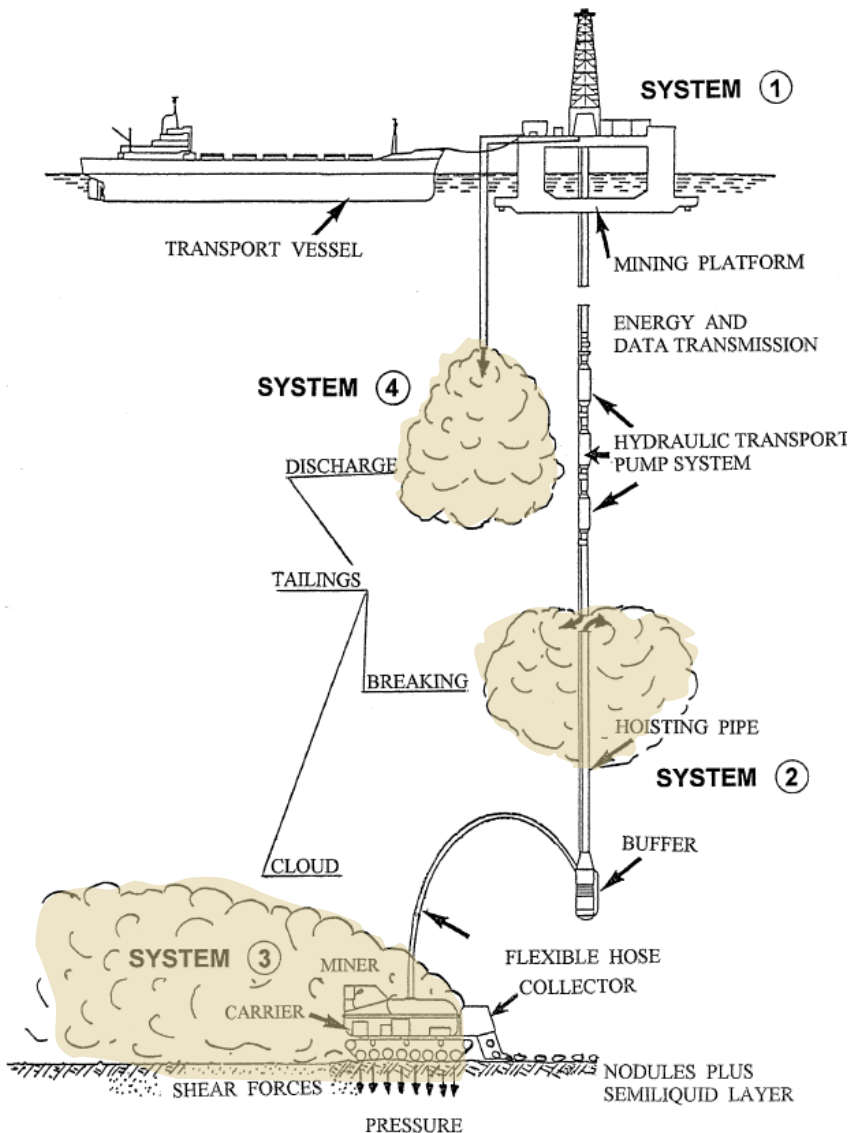
Belgian DEME-Group plans to test a nodule collector prototype (CCZ) in autumn/spring 2020/21.

Ministry of Earth Sciences (Government of India) plans technical trials for a nodule collector pre-prototype (CIOB) during 2021

Canadian Company DeepGreen plans a collector test in 2022 (CCZ)

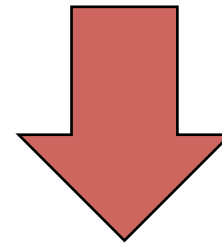
Manganese Nodules ■
Massive Sulfides ■
Cobalt-Rich Crusts ■

Impacts of polymetallic nodule mining



Physical Impacts

- Removal of nodules & 10 cm of seafloor
- Generation of sediment plume that will resettle & blanket the seafloor
- Discharge of sediment waste from surface platform / riser pipe



Potential effect on ecosystems

- Loss of habitat integrity
- Loss of species & genetic diversity
- Loss of ecosystem structure & functions

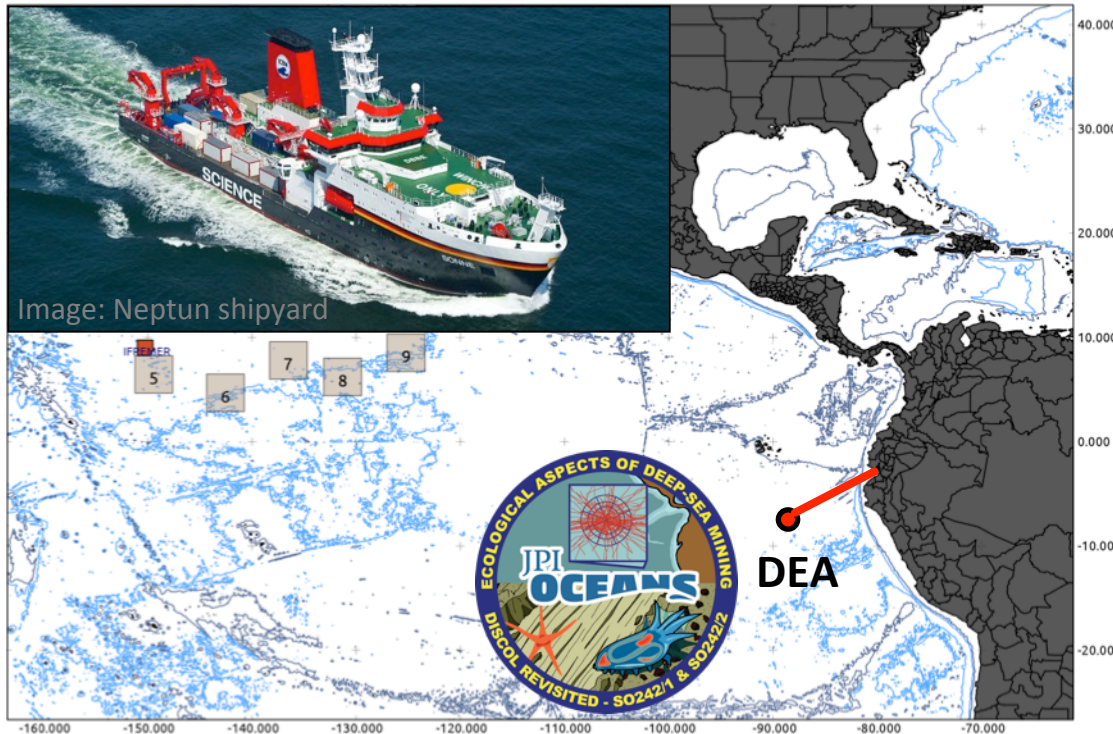
JPIO MiningImpact (2015-2018)

Overall Goal:

Assessing the long-term impacts of nodule mining on deep-sea ecosystems

Motivation and study focus

- Observations of microbial communities and functions are missing in assessments of deep-sea mining impacts

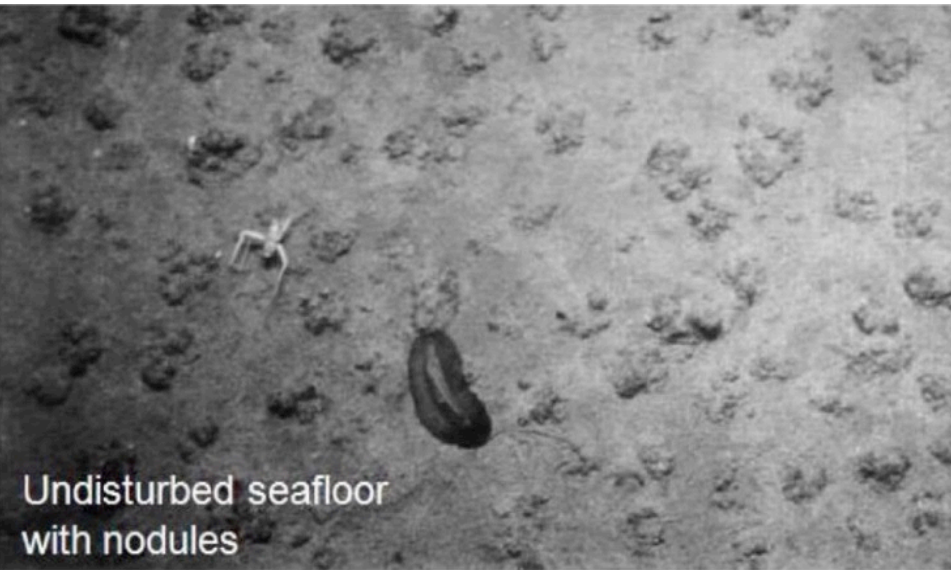


Key research questions

- can we identify disturbance effects on microbial community structure, activities, and biogeochemical functions?
- are effects scaling with disturbance intensity?
- at what time scales are communities and functions recovering?

Sampling Strategy

Comparing areas in/outside 26y old plough tracks to reference areas outside DEA



Undisturbed seafloor
with nodules

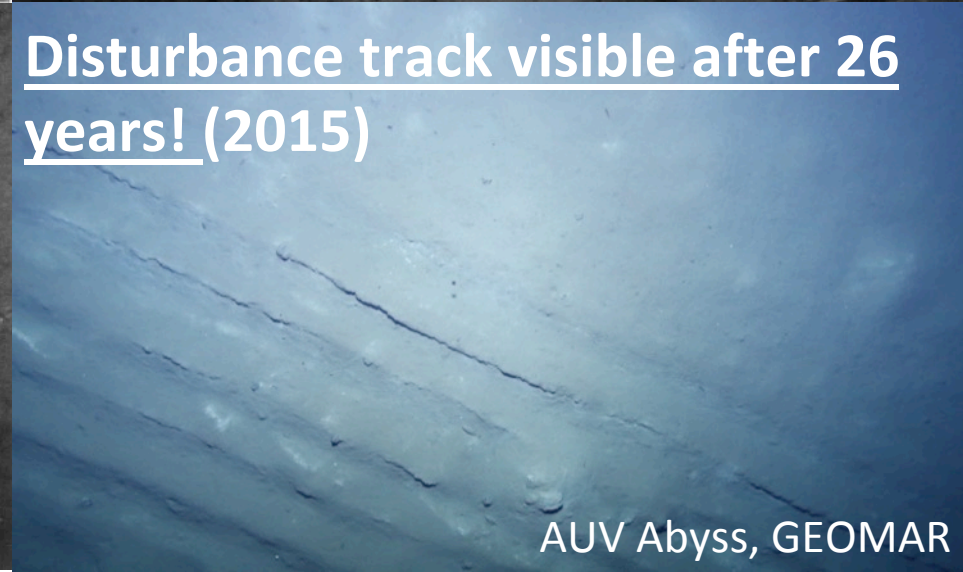


Disturbance track (0 years)



Disturbance track (7 years)

Thiel et al. (2001) DSR II 48

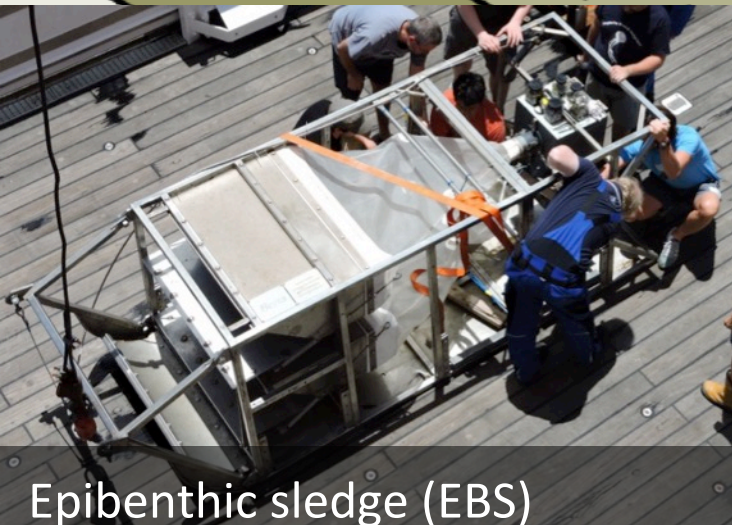
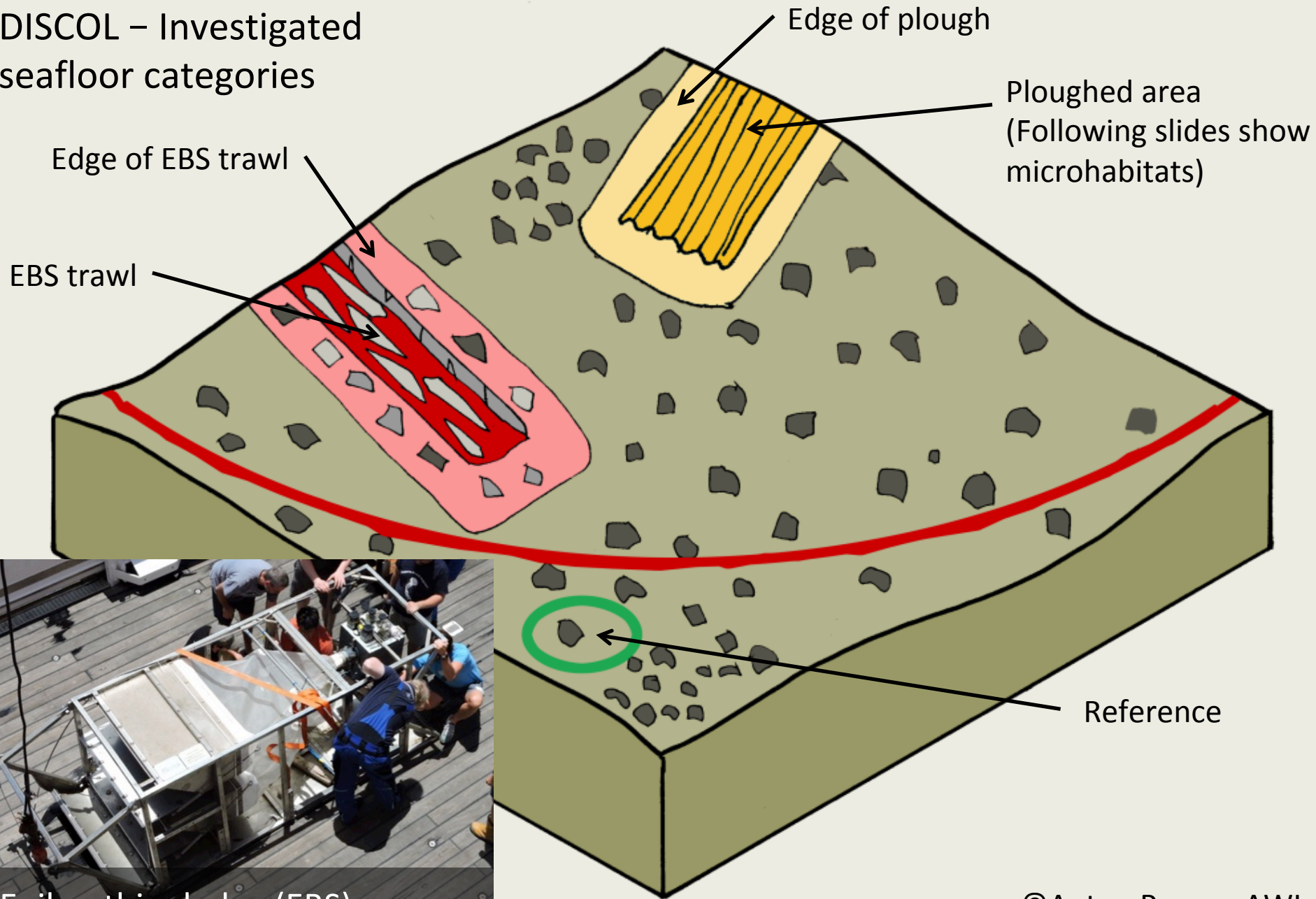


Disturbance track visible after 26
years! (2015)

AUV Abyss, GEOMAR

Sampling Strategy

DISCOL – Investigated seafloor categories



Epibenthic sledge (EBS)

Identified disturbance area microhabitats

- **Furrows**

- > surface sediments redistributed (removed?) upon ploughing

- **Ridges**

- > surface sediments redistributed (piled up?) upon ploughing

- **Subsurface patch**

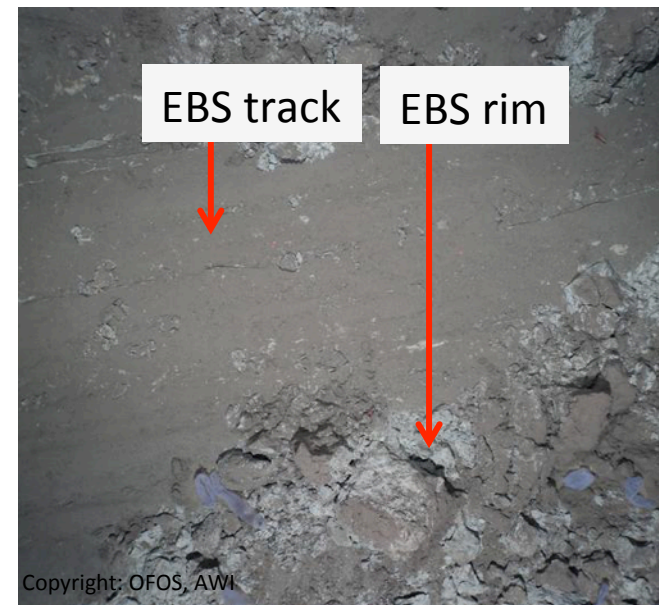
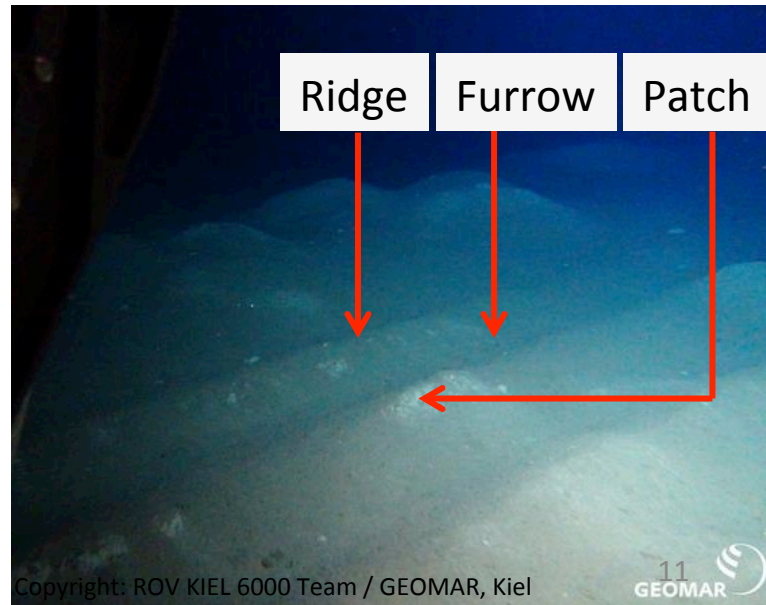
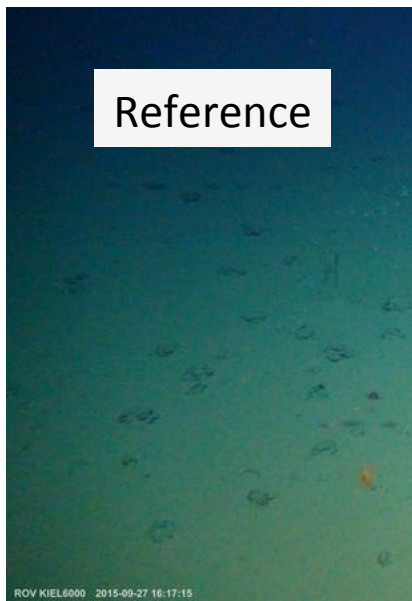
- > deeper sediment exposed on sediment surface

- **Epibenthic trawl (EBS) track**

- > fresh disturbance created during first leg (5 weeks before sampling)

DEA: plough harrow

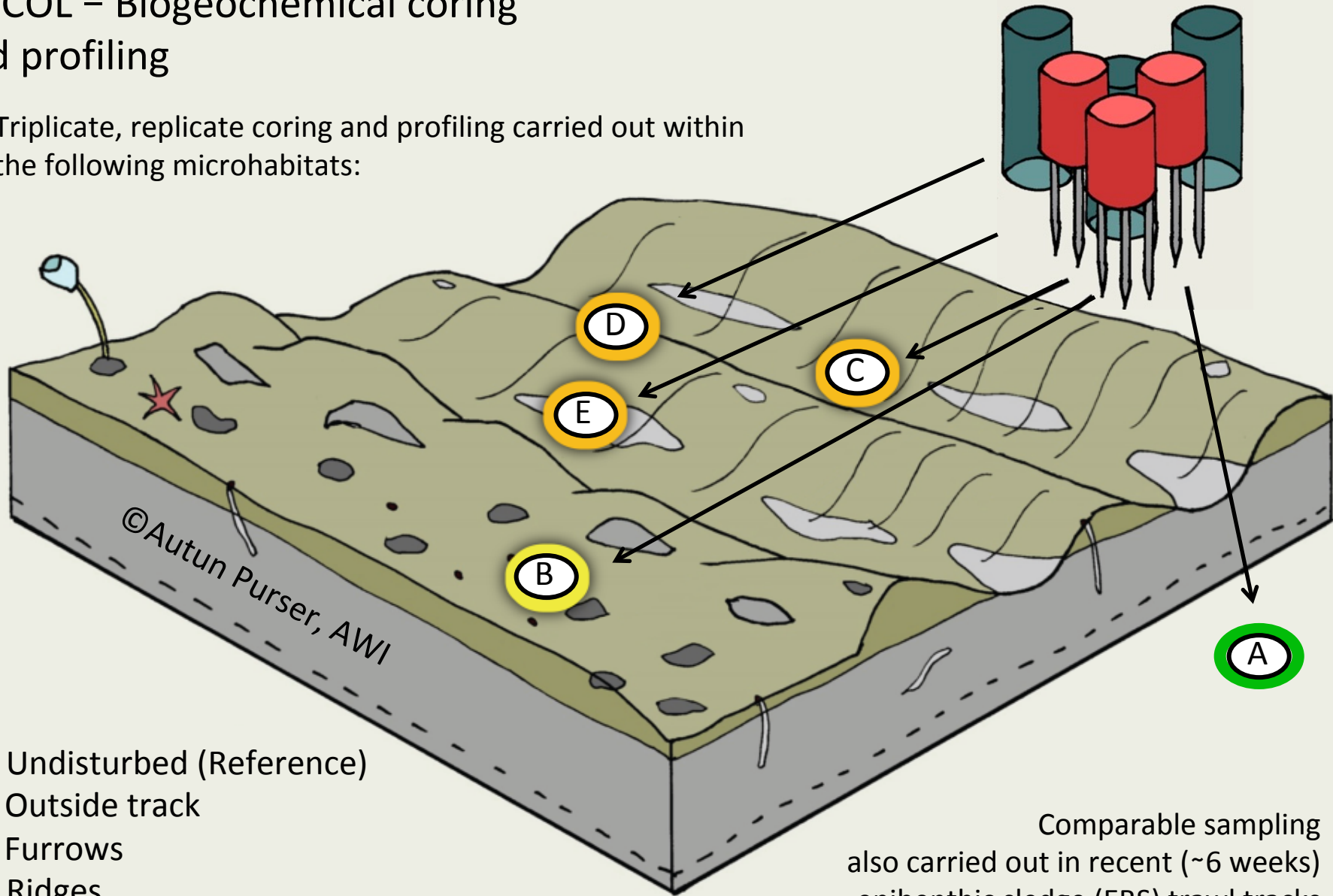
EBS



Sampling Strategy

DISCOL – Biogeochemical coring and profiling

Triplicate, replicate coring and profiling carried out within the following microhabitats:



A – Undisturbed (Reference)

B – Outside track

C – Furrows

D – Ridges

E – Subsurface patch

Comparable sampling also carried out in recent (~6 weeks) epibenthic sledge (EBS) trawl tracks made by SO242/1

Sediment biogeochemistry: quantify / classify impact intensities

Microbial abundance in Sediments:

Total cell counts (AODC)

Microbial community structure and diversity in Sediments & Nodules:

Illumina sequencing of 16S rRNA gene:

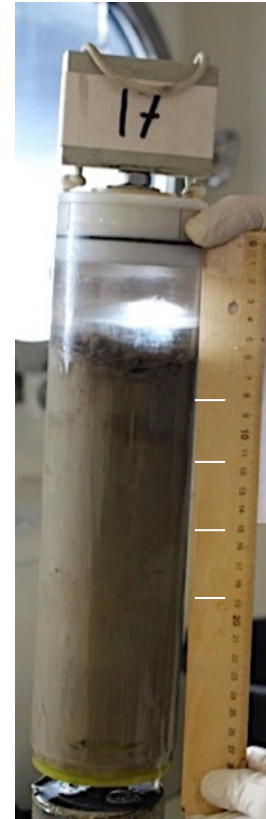
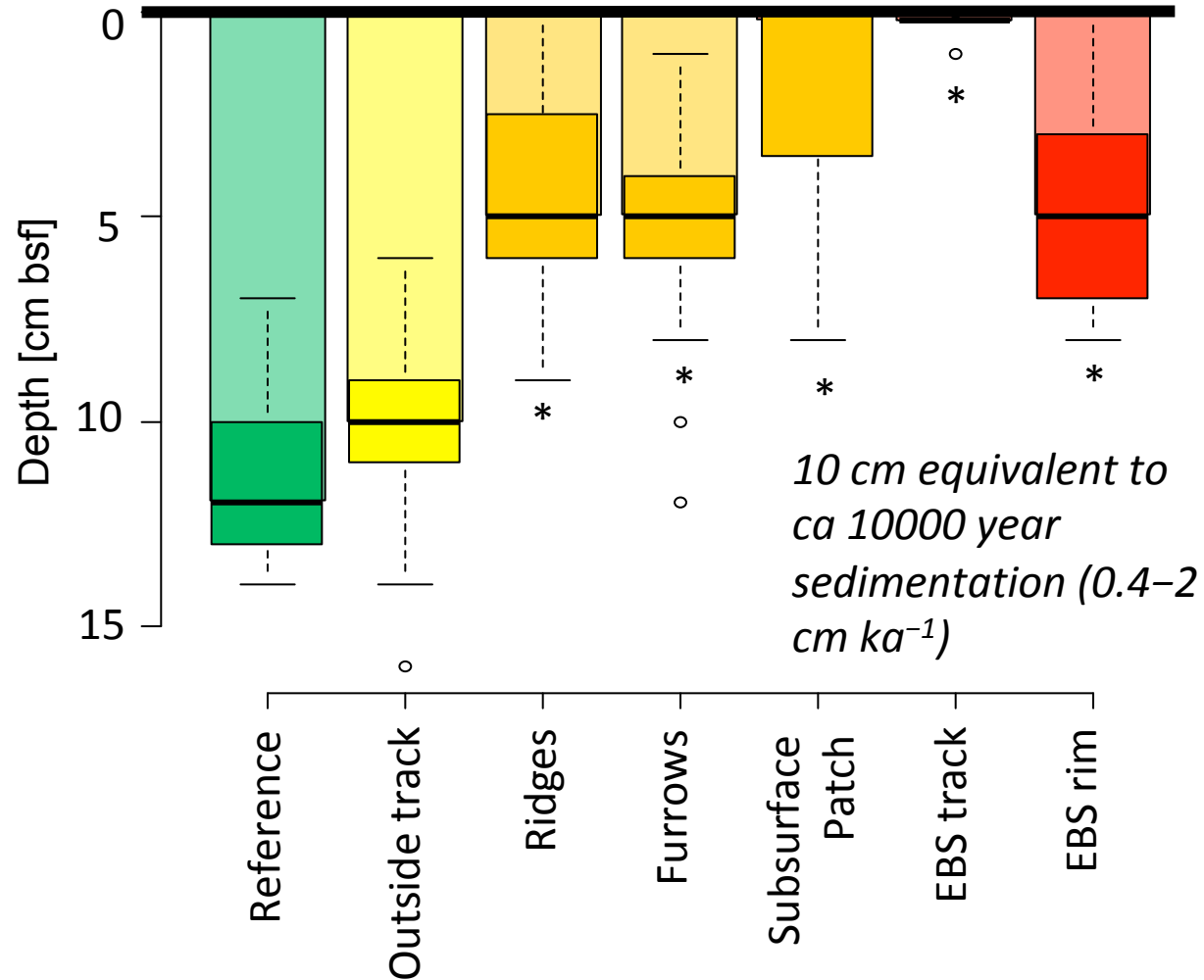
Bacteria V3-V4 region

Archaea V3-V5 region

Microbial activity in Sediments

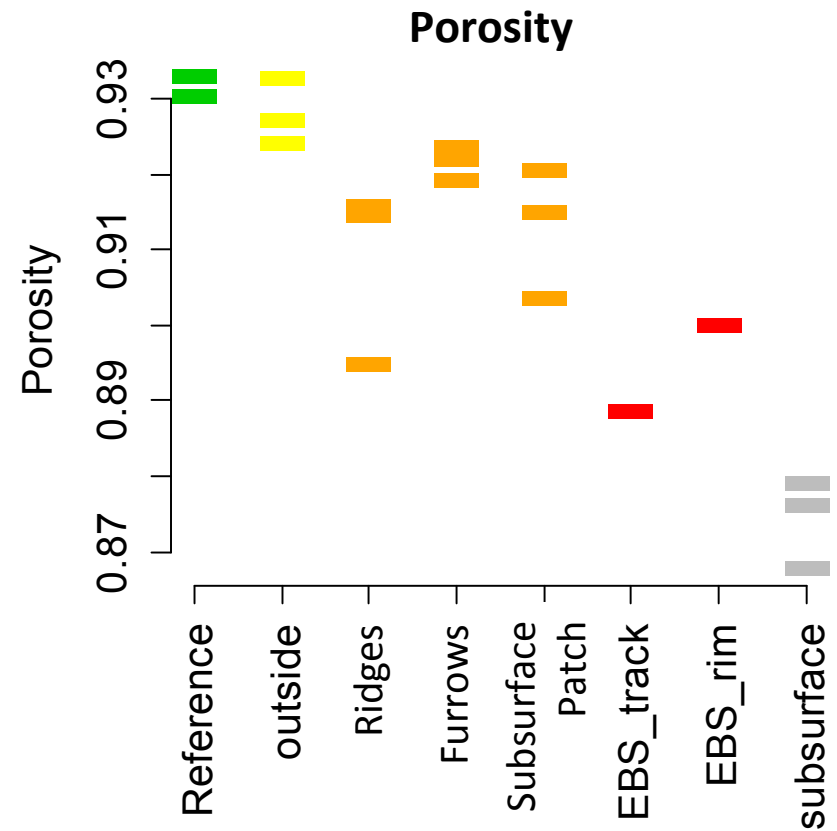
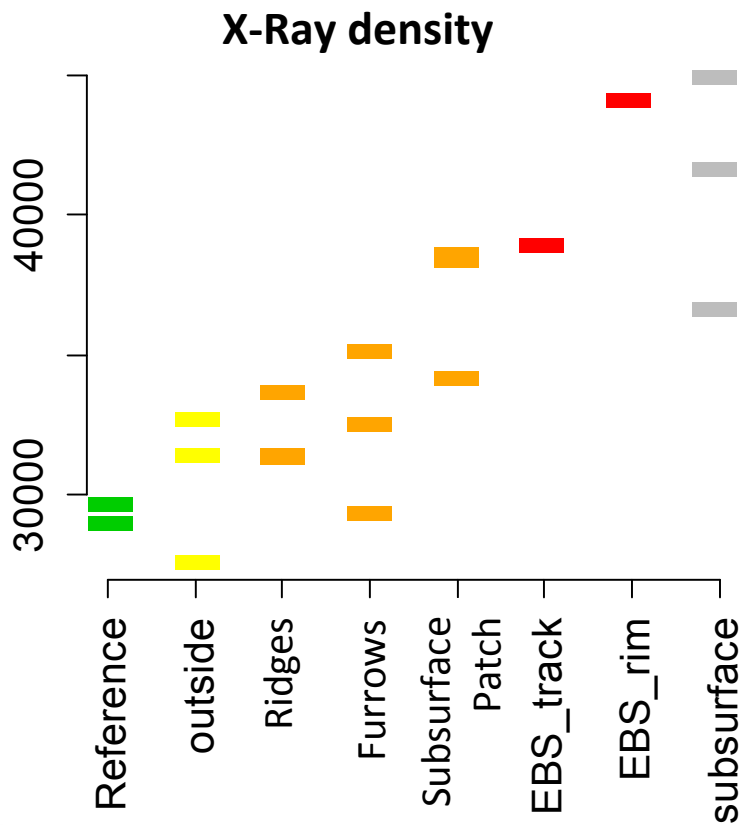
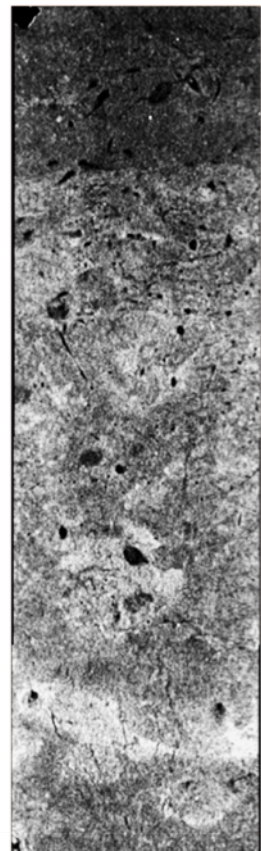
- Potential OM degradation: extracellular enzymatic activity (EEA)
- OM remineralization: in situ O₂ microsensor sediment profiles and chamber incubations
- Substrate uptake rates: ¹⁴C-bicarbonate (DIC) and ³H-leucine
- Biomass production: empirical DIC and leucine to C conversion factors

- Disturbance & loss of reactive surface layer



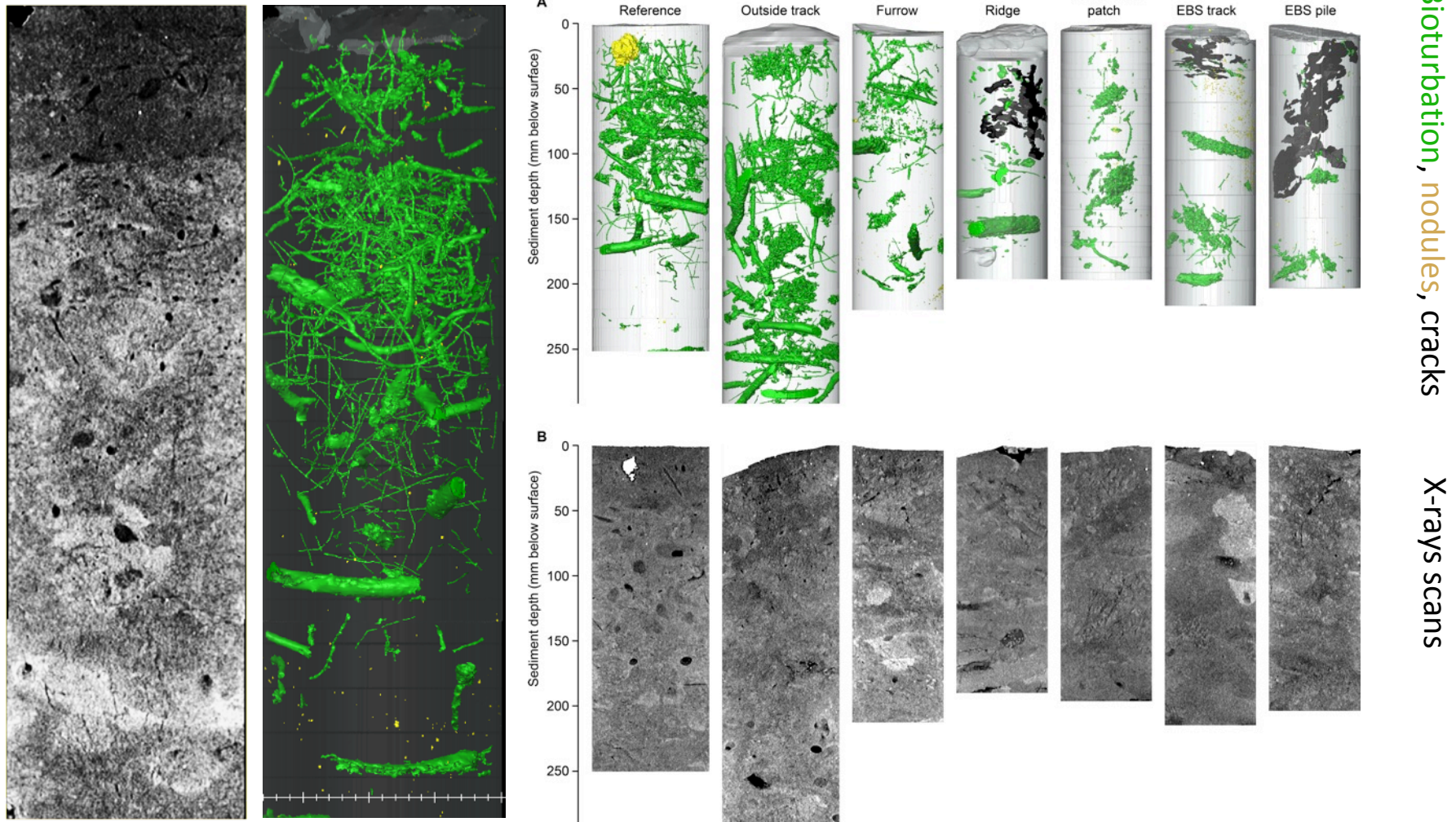
Changes in seafloor properties due to ploughing

- Stiffer, more compacted sediment exposed => difficult to recolonize => low bioturbation activity



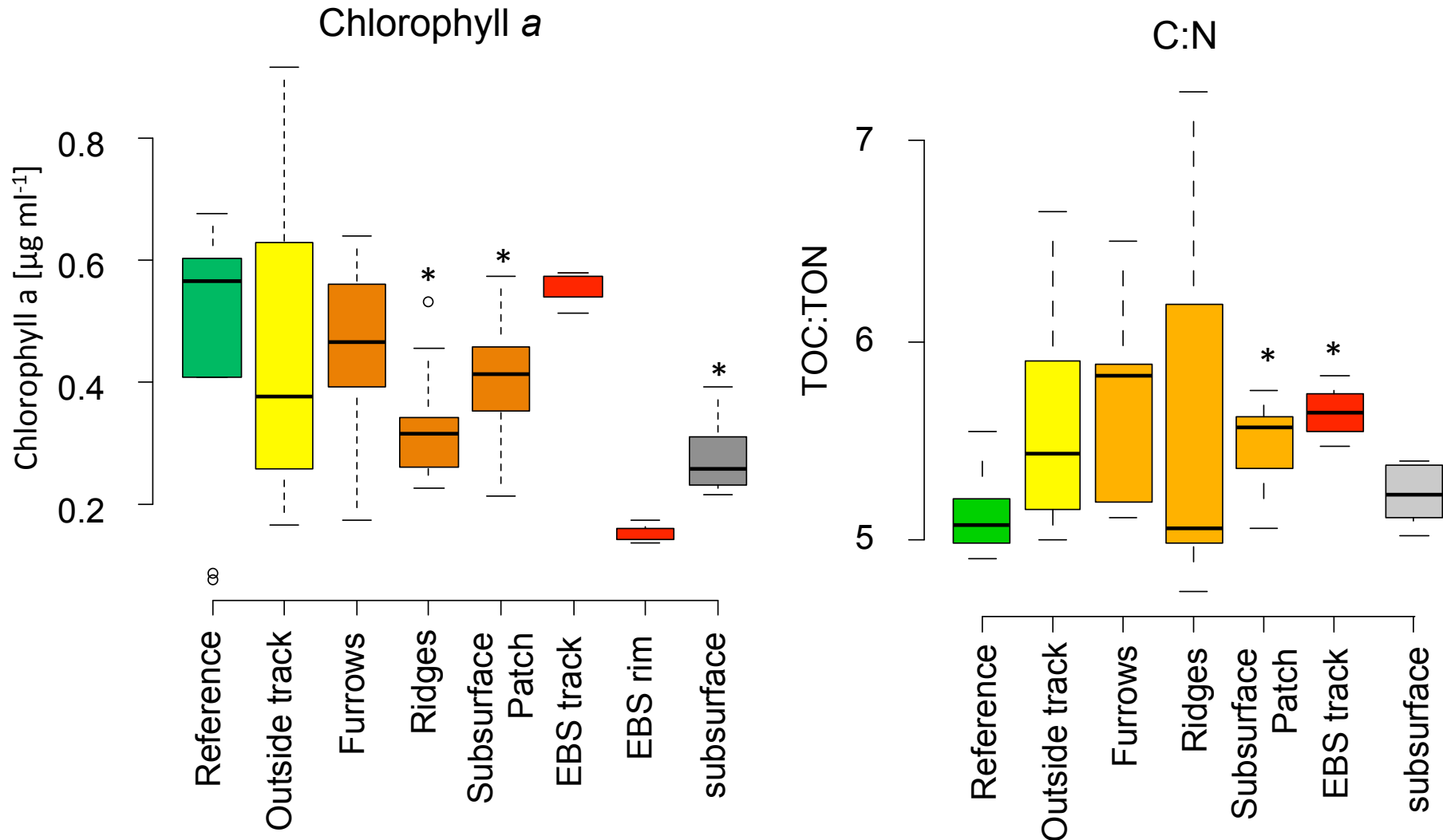
Impacts on bioturbation (DEA)

- X-rays show no / little recovery of bioturbation activity in 26 year old tracks
- exposed stiffer subsurface sediments difficult to colonize

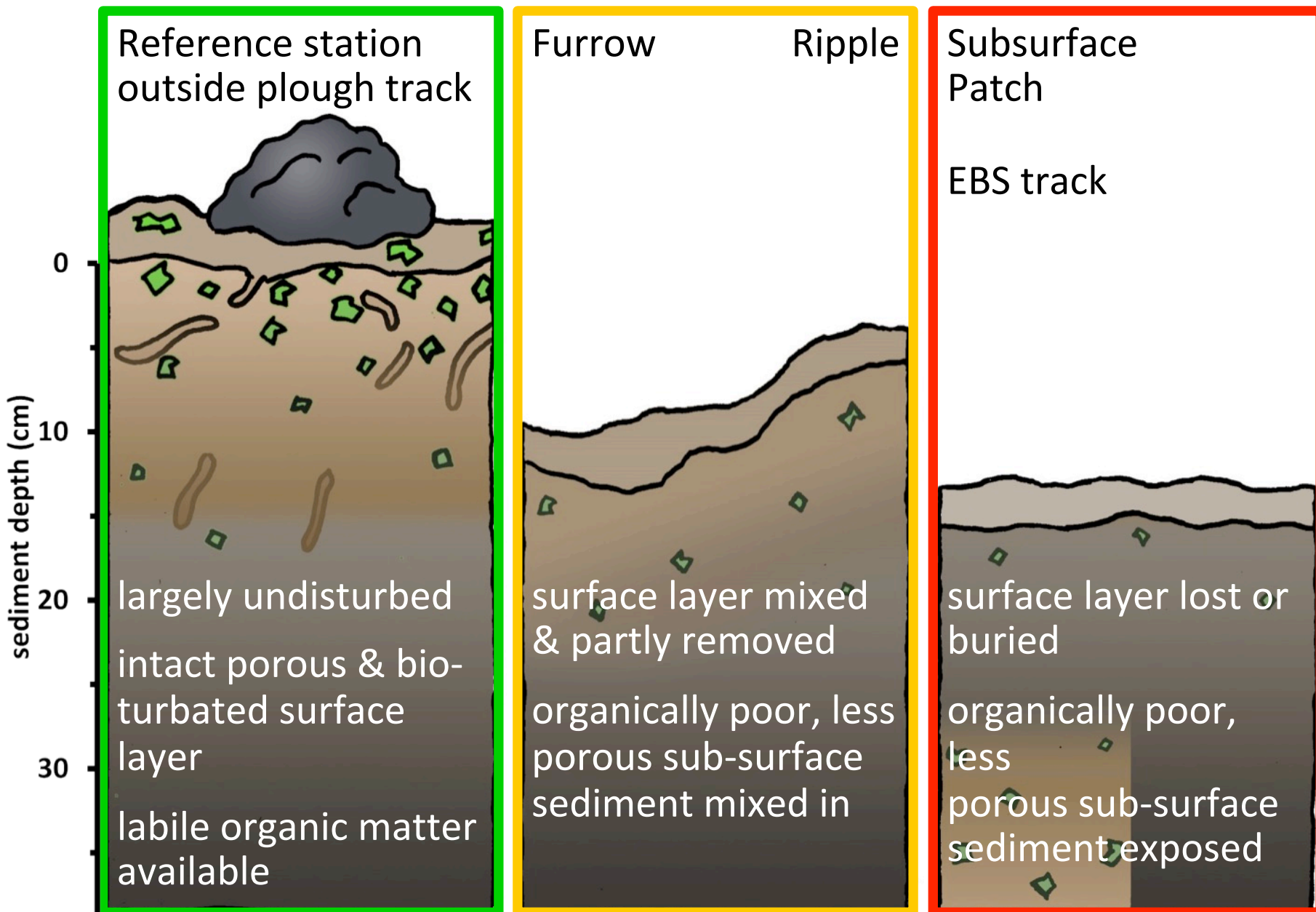


Loss of labile organic matter (DEA)

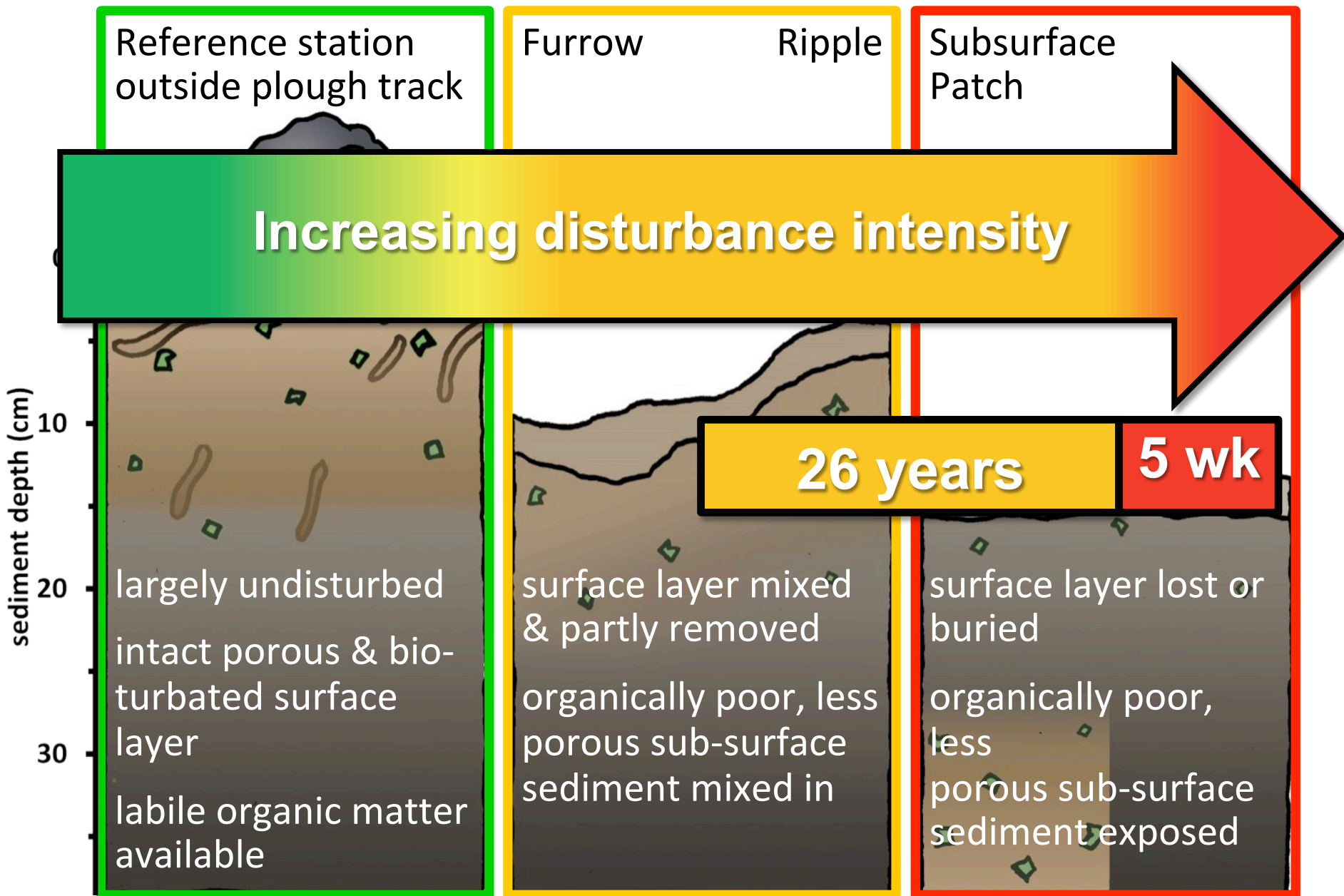
- Fresh detritus & nitrogen-rich organic matter is still reduced 26 y after disturbance



Disturbance intensity gradient scheme (Vonnahme et al. 2020, Science Advances)

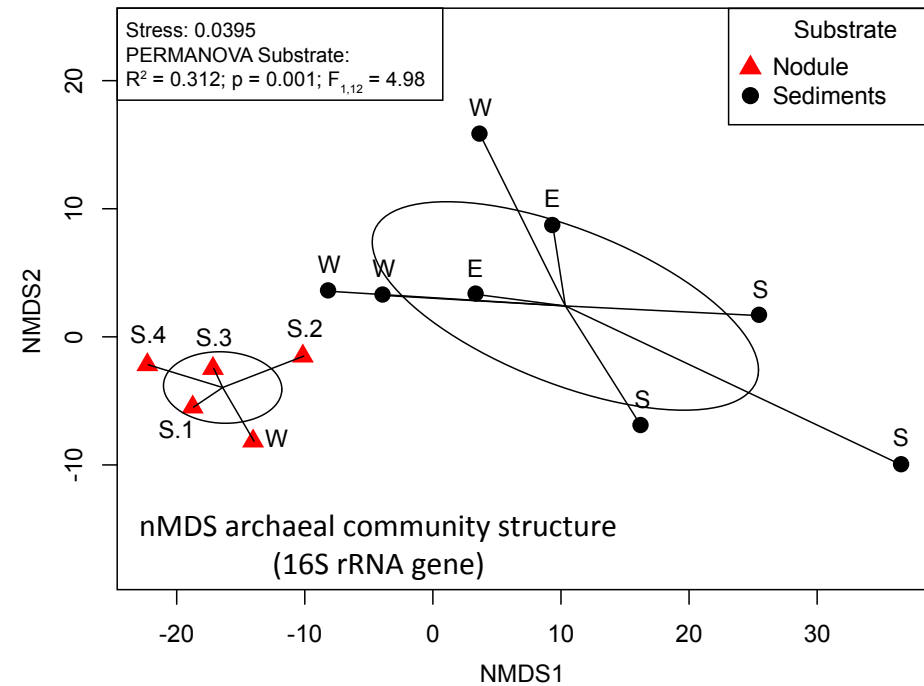
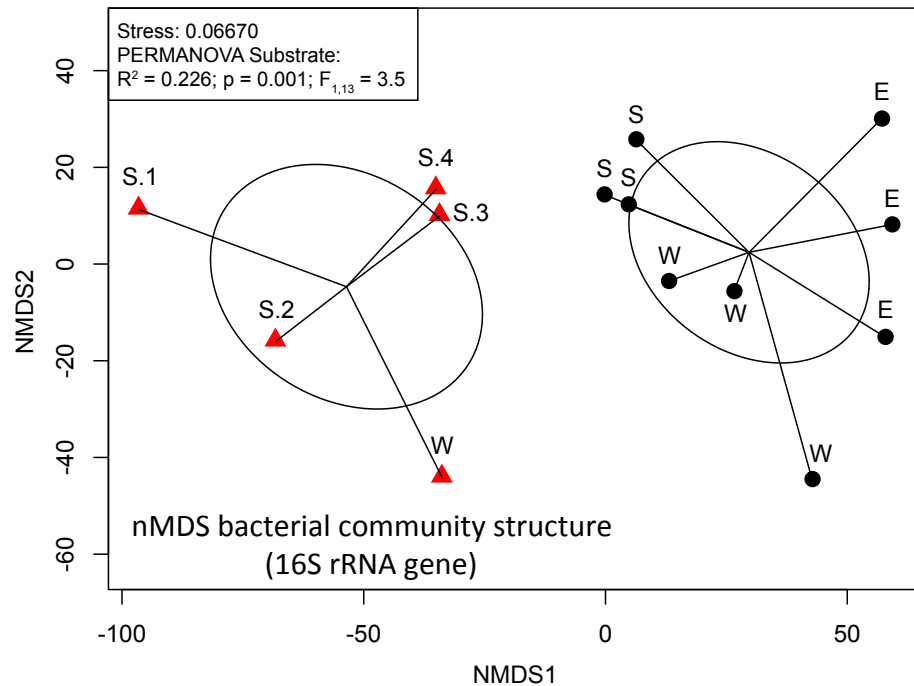


Disturbance intensity gradient scheme (Vonnahme et al. 2020, Science Advances)



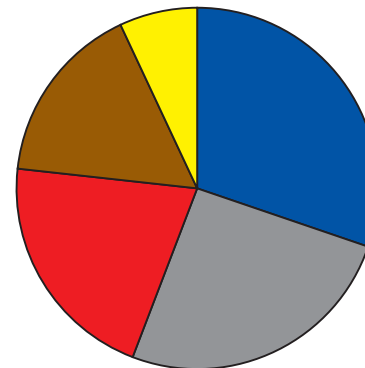
Results | Impact on bacterial diversity and community by nodules removal

- Nodules and sediments host distinct bacterial and archaeal communities



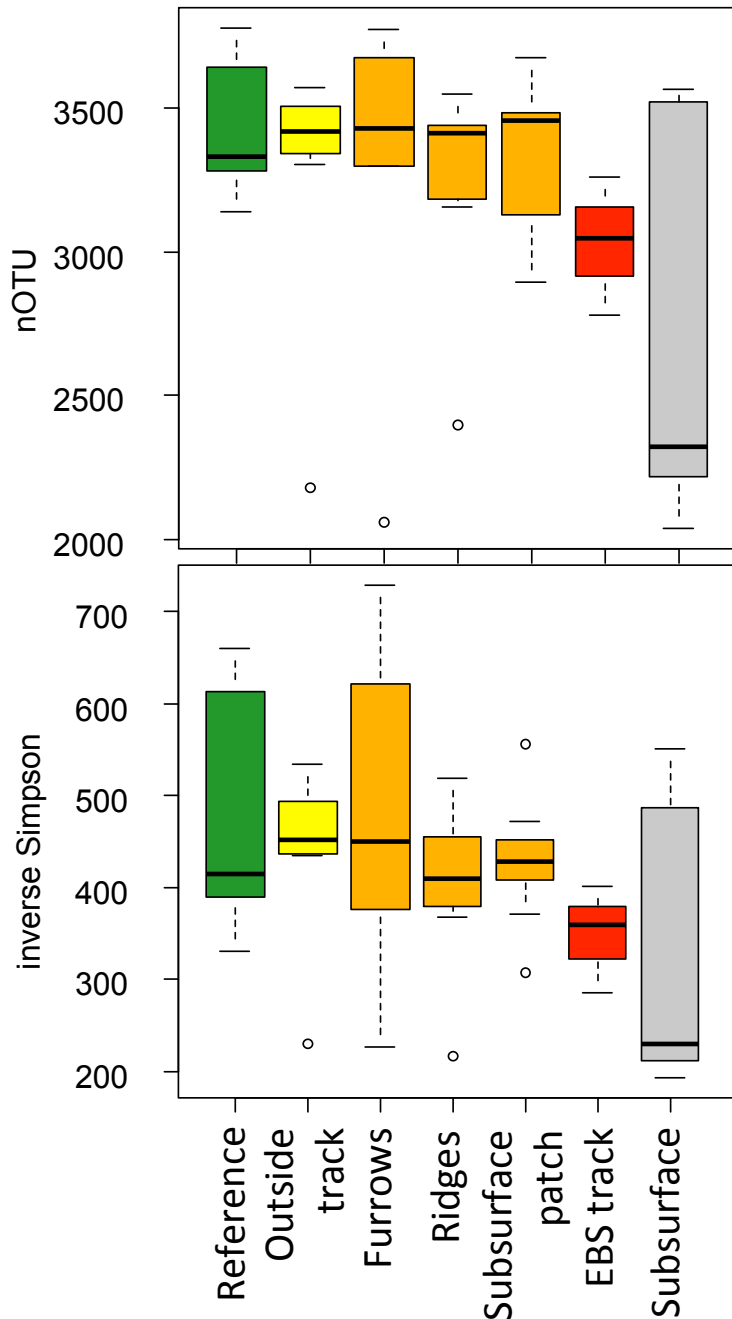
- Nodules are enriched in:

- potential metal-cycling bacteria (i.e. Magnetospiraceae, Hyphomicrobiaceae)
- bacterial and archaeal nitrifiers (i.e. *AqS1*, unclassified Nitrosomonadaceae, *Nitrosopumilus*, *Nitrospina*, *Nitrospira*)
- unclassified bacterial sequences found in ocean crust, others nodule fields, hydrothermal deposits and sessile fauna.

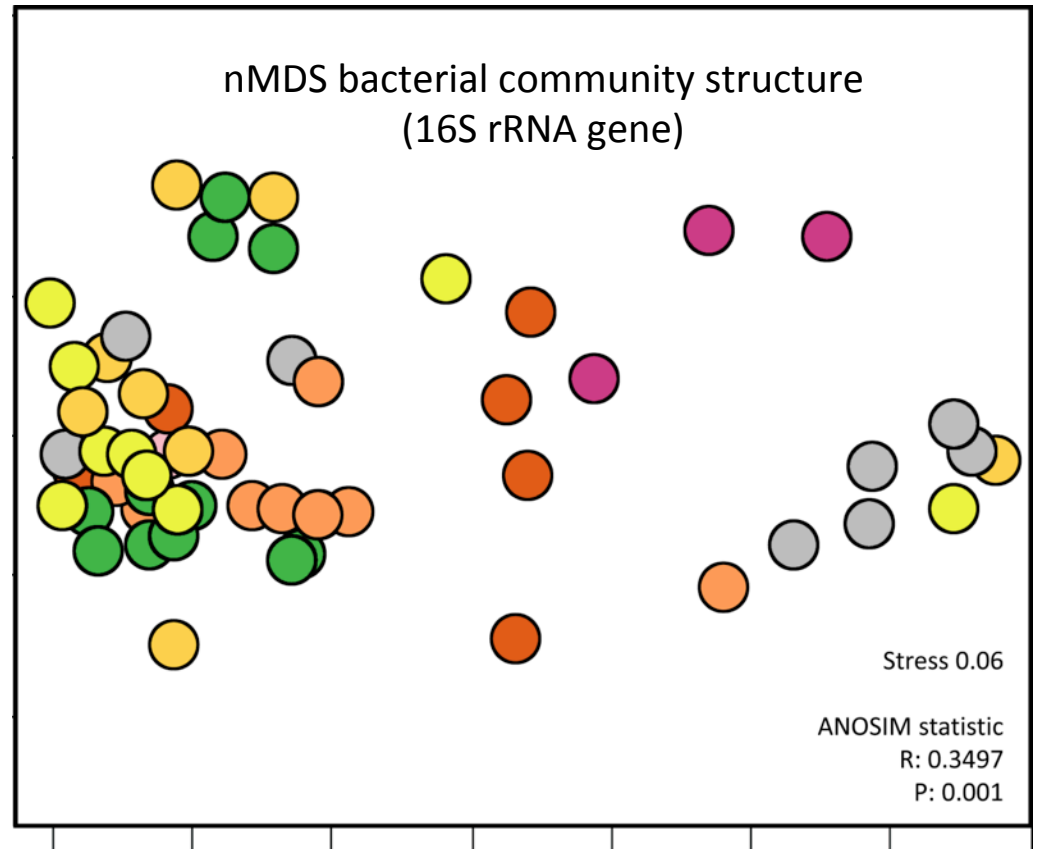


- Ocean crust
- Nodule field and nodule
- Hydrothermal/Seep sediments and deposits
- Deep-sea sediments
- Invertebrates

Results | Impact on bacterial diversity and community

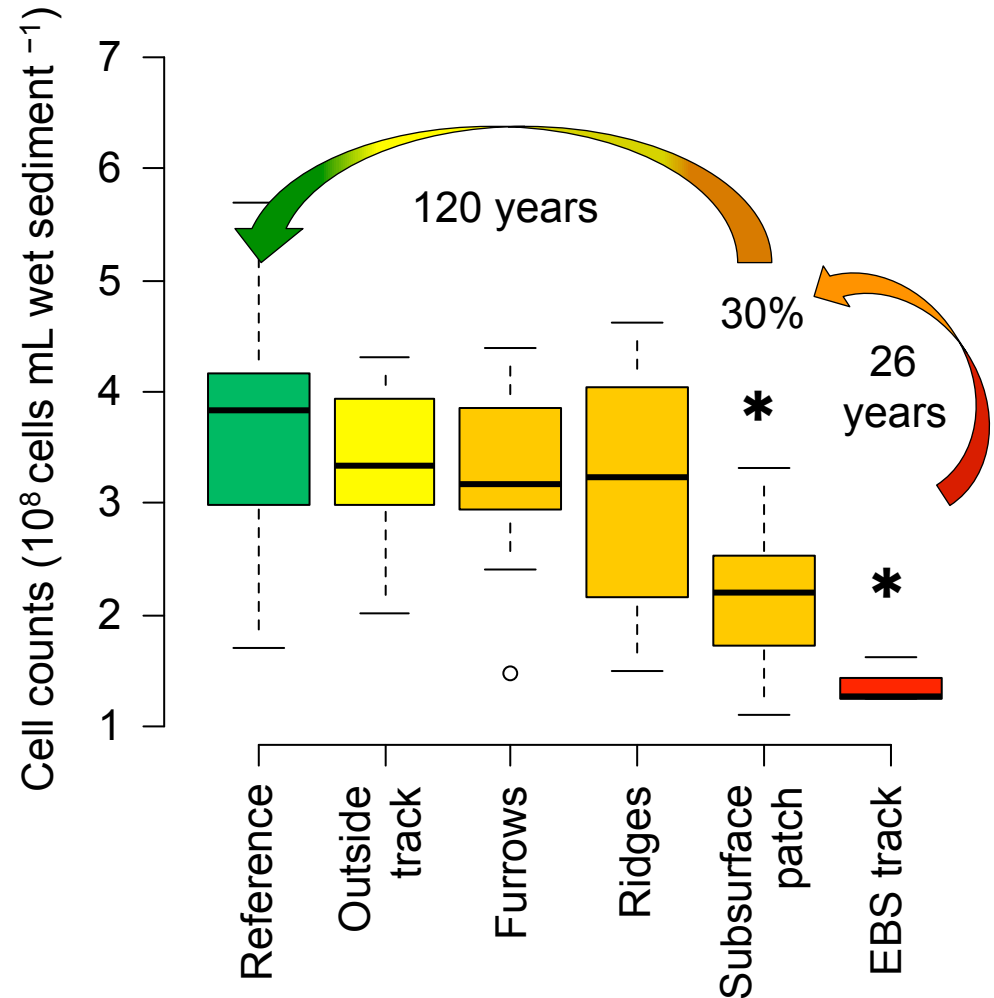
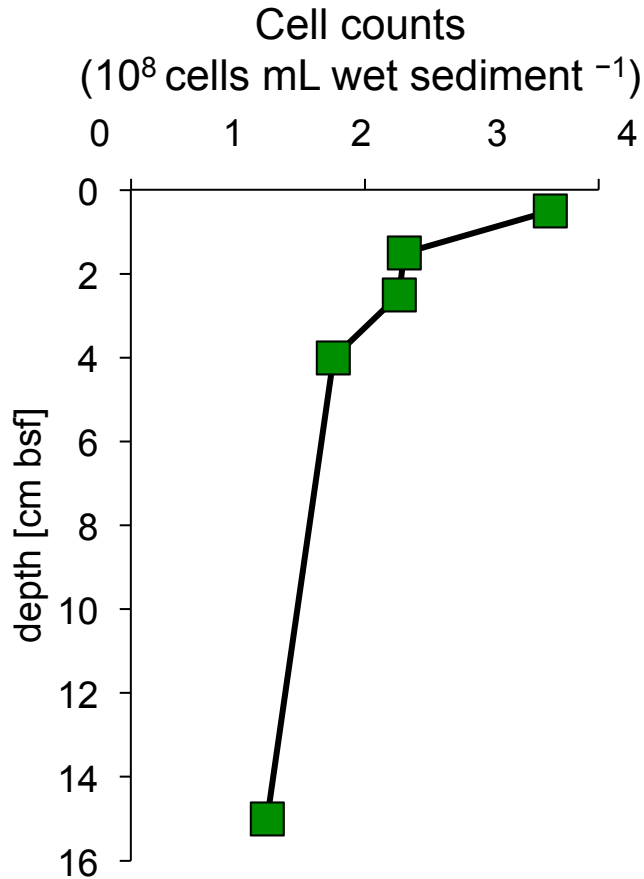


- Indications of disturbance related changes in microbial communities, but effects are hard to discriminate from pronounced spatial variability



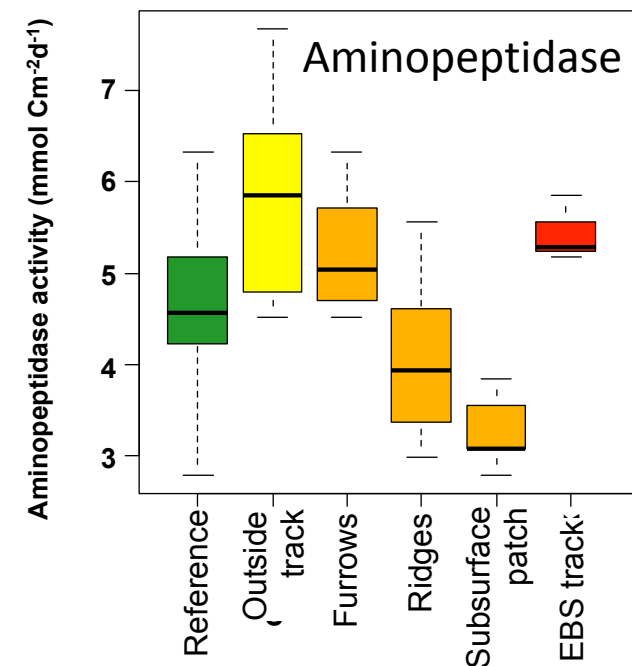
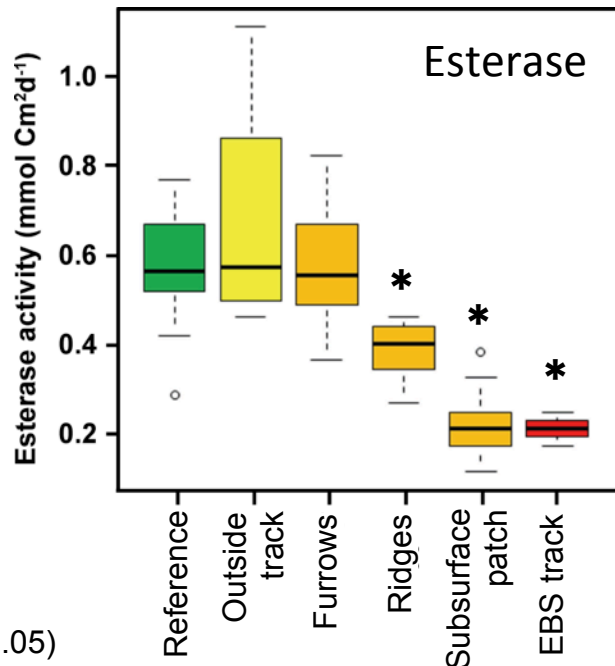
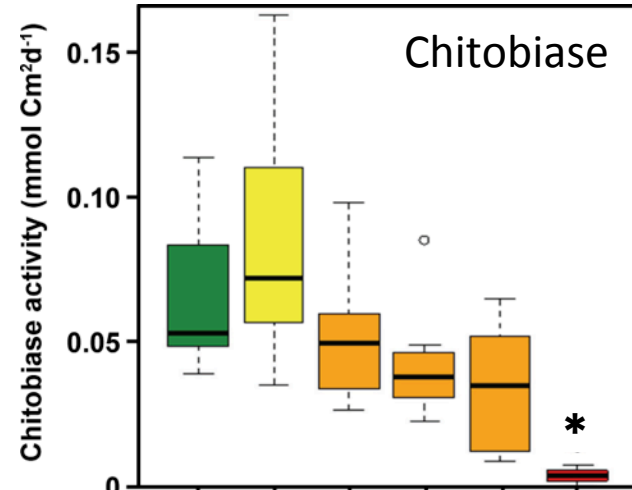
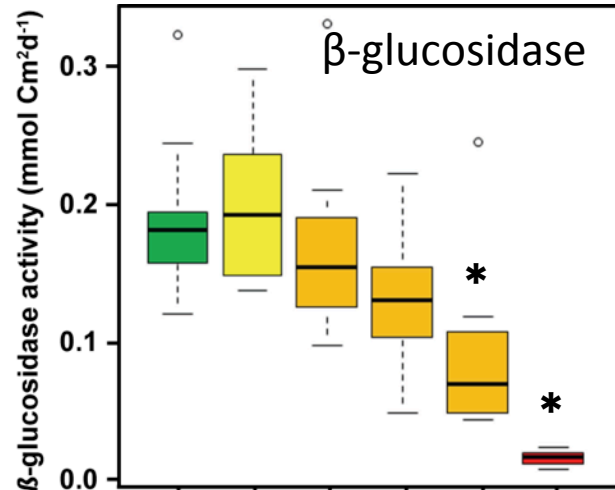
Results | Impact on microbial standing stock and activity

- Cell abundances are still reduced after 26 years in some microhabitats
- Recovery time estimated to be ~ 100 years



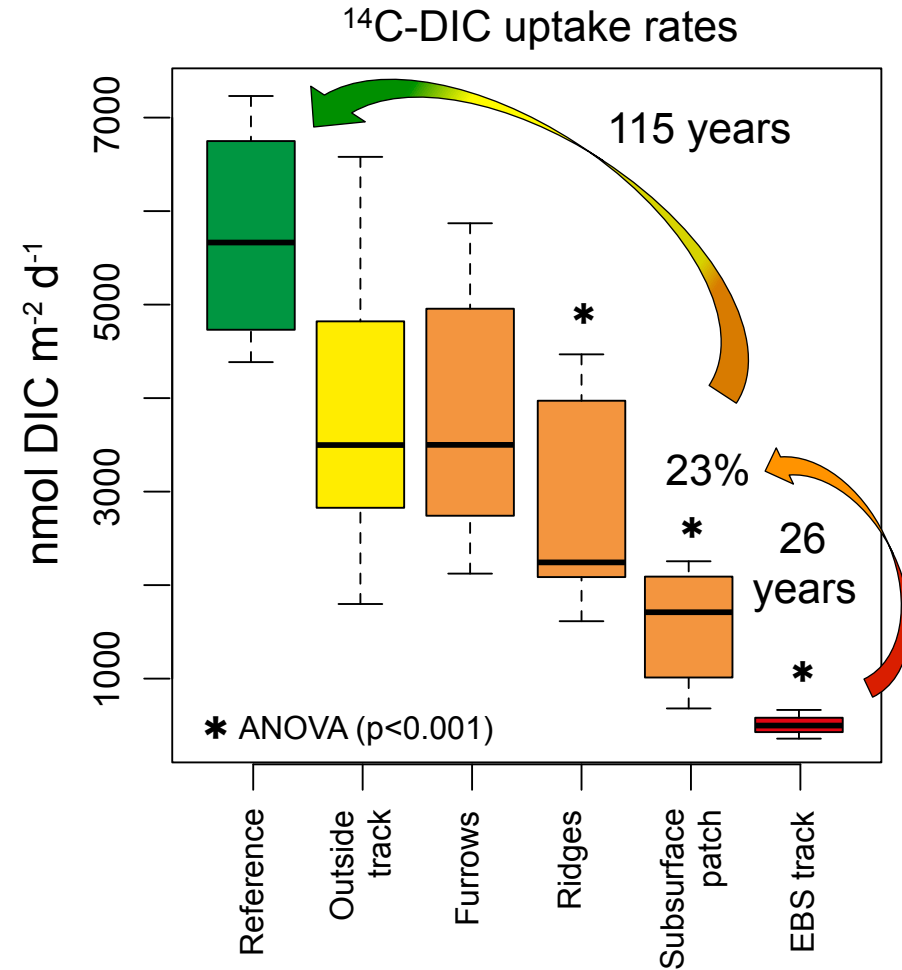
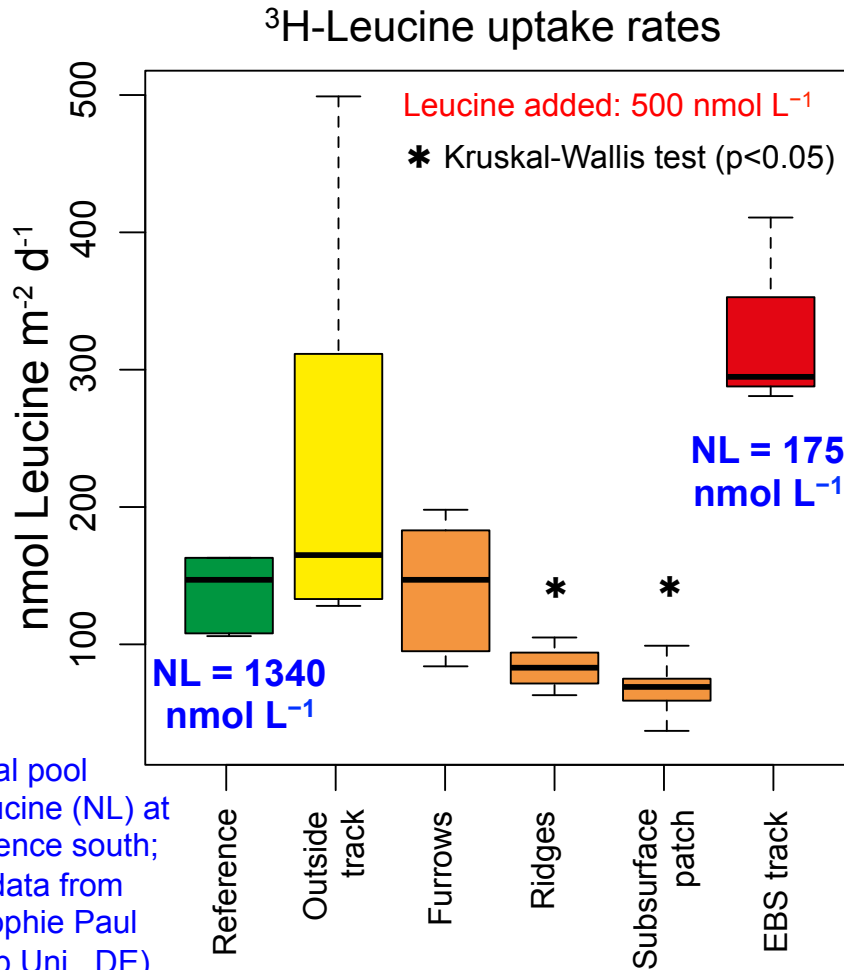
Results | Impact on microbial standing stock and activity

- Strong effects on extracellular enzymatic activities in disturbance tracks (especially where surface sediments are buried or lost)



Results | Impact on microbial standing stock and activity

- Strong effects also on biomass production (inorganic C and Leucine uptake)
- Recovery time estimated to be ~ 100 years
- Stimulation of Leucine uptake in EBS track suggests food limitation



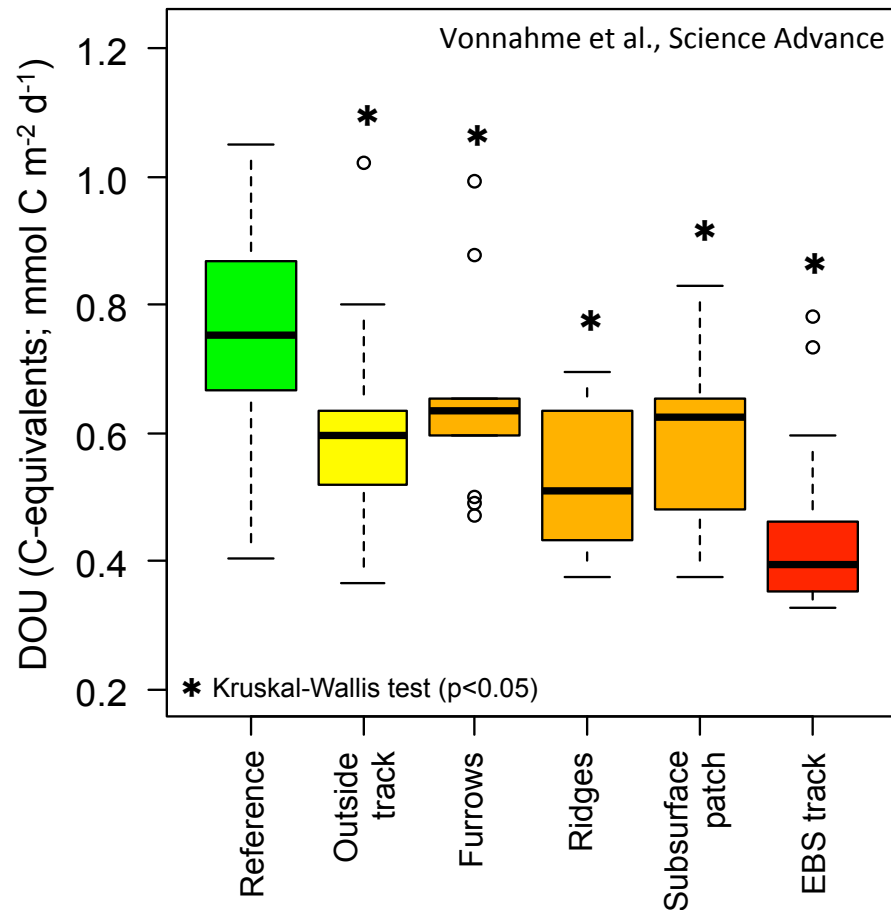
Natural pool of Leucine (NL) at Reference south; DAA data from Dr. Sophie Paul (Jacob Uni., DE)

Natural pool of leucine (NL) assessed with saturation curves at Reference East: 1150-1375 nmol/L

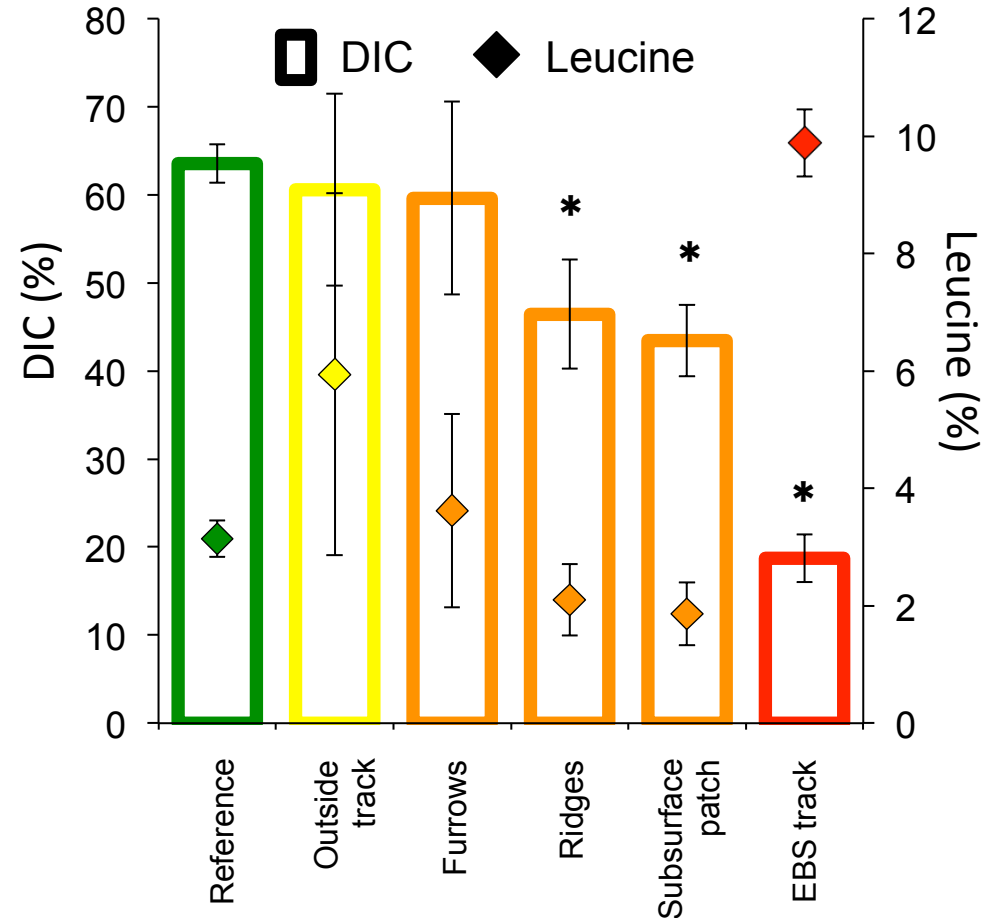
Results | Impact on microbial standing stock and activity

- Oxygen fluxes and grow efficiency generally decreasing with increasing disturbance intensity

O₂ respiration



Grow efficiency [BP/(BP+R)]



Empirical Leucine CF = 1.90 Kg C mol⁻¹ Leu (this study, unpublished data)

[theoretical CF: 1.44 -1.55 Kg C mol⁻¹ Leu; Simon and Azam (1989); Buesing and Marxsen (2004)]

Empirical DIC CF = 2.54 Kg C mol⁻¹ DIC (this study, unpublished data)

Conclusions

- Ploughing created different microhabitats with characteristic changes in physical and biogeochemical conditions
- High spatial variability in microbial communities and activity require extensive baseline studies
 - > *Reference / conservation areas need to match characteristics of mined areas (e.g. productivity, nodule coverage, topography) and replication during baseline studies and monitoring needs to cover local variability*
- Carbon cycle (OM degradation and remineralization, and C transfer to food web) is highly impacted and did not fully recover in 26 years (estimate time ca. 100 years)
 - > *Identification of key active microbial taxa and processes would help monitoring the recovery of the ecosystem*
- Nodules represent a specific ecological niche (i.e. hard substrate, high metal concentrations, and sessile fauna), with a potentially relevant role in organic carbon degradation and in the cycling of elements.
 - > *Assessment metabolic activities / Restoration experiments*

Acknowledgements



Max Planck Institute
for Marine Microbiology



MAX-PLANCK-GESELLSCHAFT

- JPI Ocean project 'Ecological Aspects of Deep-Sea Mining'
- Germany's Federal Ministry of Education and Research for national funding within JPIO & RV SONNE expedition costs
- Max Planck Society & Helmholtz Society for additional funds
- Captain and crew of RV SONNE & JPIO 'MiningImpact' project partners for support on board

Autun Purser	Halina Tegetmeyer
Yann Marcon	Jana Bäger
Jakob Barz	Johannes Lemburg
Axel Nordhausen	Erika Weiz
Fabian Schramm	Rafael Stiens
Mirja Meiners	Martina Alisch
Viola Beler	Cäcilia Wigand

00_2015-04-14 22:01:06

GEOMAR 

 **HELMHOLTZ**
ASSOCIATION

JPI
OCEANS



Federal Ministry
of Education
and Research